

Original Research Article

Characterization of Hepatitis C Virus (HCV) Genotypes with Spectrum of its Viral Load

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Abstract: The Hepatitis C virus (HCV) is the most common cause of chronic liver disease and is increasing worldwide. Proposed study includes 24 clinical samples, processed for HCV RNA viral load and HCV genotyping. HCV RNA viral load ranges in between 1.00×10^1 IU/ml to $> 1.00 \times 10^8$ IU/ml and the cases with high viral load were further processed for the genotyping. Out of 24 specimens characterized genotype 3a was prevalent in 72.72% (8/24) cases and HCV genotype 1b was in 27.27% (3/24) cases followed by HCV genotype 4a, which was in 27.27% (3/24) cases. In 13 cases target was not detected due to low viral load. 8 cases were for HCV genotype 3a of which 5 were males & 3 females, followed by HCV genotype 1b (3 males) & 4a (3 females). The genotype 3a was found in 8 (72.72%) patients. Genotype 1b was seen in 3 (27.27%) patients. Genotype 4a was seen in 3 (27.27%) patients. Genotype 2a was seen in only 1 (9.09%) patient. From the current study genotype 3a was most prevalent and can be further studied. The outcome of HCV genotyping is of almost clinical value as there are various regimens available to treat different types of HCV genotypes.

Keywords: Genotype, Sexual transmission, cirrhosis, Real Time PCR, replicating viruses, Chronic.

INTRODUCTION:

Hepatitis C virus (HCV) is an enveloped single-stranded RNA virus which appears to be distantly related to flaviviruses, although HCV is not transmitted by arthropod vectors [1, 2]. Hepatitis C virus is associated with chronic liver disease and also with primary liver cancer in many countries. HCV is mostly transmitted through exposure to infective blood [3]. This may happen through transfusions of HCV-contaminated blood and blood products, contaminated injections during medical procedures, and through injection drug use [4, 6]. Sexual transmission is also possible, but is much less common. Unless successfully treated with medication, chronic Hepatitis C infection can cause other serious health problems, such as cirrhosis, liver cancer and liver failure [7, 10]. Hepatitis C is divided into six distinct genotypes throughout the world with multiple subtypes in each genotype class. In the current project the patient with HCV infection were considered & further Quantification of 5'untranslated region of the HCV genome & its genotypic

characterization were determined in spectrum of its viral load [11,15].

MATERIALS AND METHODS:

A total of 24 clinical samples were considered in this study. Blood samples were taken from the infected cases of HCV and further processed at Central Molecular Research Laboratory (CMRL), Shri Guru Ram Rai Institute of Medical & Health science (SGRRIM & HS), Patel Nagar, and Dehradun (U.K.) for the quantification of HCV & identification of HCV genotypes. Pre-processing of the sample collection: 5 ml EDTA Whole blood was centrifuged at 5,000 rpm for 10-15 minutes. Isolated serum was used further for RNA isolation. RNA was isolated by High Pure Viral Nucleic Acid Extraction kit. The specimens were processed for HCV viral load by Roche COBAS TaqMan 48 analyzer & the 5'untranslated region of the HCV genome was amplified for the quantification of HCV virus [16, 17]. Further HCV genotypic characterization was achieved by nested PCR targeting

core region with genotype specific PCR primers [18, 19].

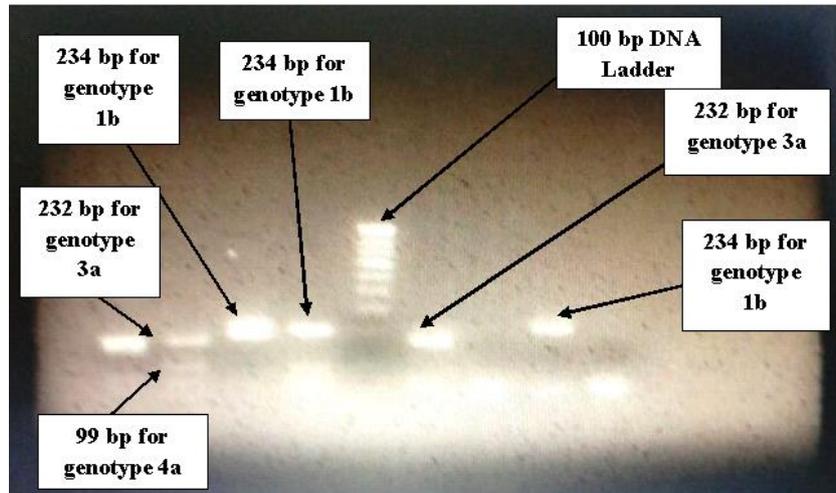


Fig 1: Gel picture showing different genotypes of HCV at different base pairs

- Well No.1- HCV genotype 3a (232 bp)
- Well No.2- HCV genotype 3a (232 bp), HCV genotype 4a (99 bp)
- Well No.3- HCV genotype 1b (234 bp)
- Well No.4- HCV genotype 1b (234 bp), HCV genotype 4a (99 bp)
- Well No.5- 100 bp DNA Ladder
- Well No.6- HCV genotype 3a (232 bp)
- Well No.7- Negative (Genotype not detected due to low viral load)
- Well No.8- HCV genotype 1b (234 bp)

RESULTS:

A total of 24 clinical samples were collected from different Departments of Shri Mahant Indiresch Hospital, Patel Nagar, Dehradun U.K which includes medicine, surgery, pediatric, gastro etc were included for the proposed study. It was seen that viral load of HCV RNA ranges in between 1.00×10^1 IU/ml to $> 1.00 \times 10^8$ IU/ml. Sample having the high viral load were further processed for the genotyping & those less than 2.0×10^3 IU/ml were not considered because the sensitivity is low of the assay. The nested PCR yielded different amplicons for different HCV genotypes (as depicted in fig. 1). Out of 24 specimens characterized genotype 3a was prevalent in 72.72% (8/24) cases and HCV genotype 1b was in 27.27% (3/24) cases followed

by HCV genotype 4a, which was in 27.27% (3/24) cases. In 13 cases target was not detected due to low viral load (limit of detection of the protocol is 25 IU/ml of HCV RNA) & all these 13 cases were not considered for further genotyping. It was also observed that in viral load ranging in between 1.00×10^4 to 1.00×10^7 IU/ml maximum number of cases were 10 and in the same spectrum HCV genotype 1b, 2a, 3a, 4a were characterized as tabulated in table no.1,2 and 3. Out of 24 samples different age groups were also considered which includes from 0-20 years, 21-40 years, 41-60 years, & above 60 years, & the number of cases in these particular age groups were 1 (Female), 8 (7 Male & 1 Female), 14 (Males), 1 (Female) respectively.

Table 1: Spectrum of HCV RNA viral load & its genotypes characterization

S.N o.	HCV RNA Titer (IU/ml)	No. of Cases	HCV Genotypes	1a	1b	2a	2b	3a	3b	4a	5a
1.	Target not detected	13									
2.	$1.00 \times 10^1 - 1.00 \times 10^3$	1	1b,3a,4a	-	1	-	-	1	-	1	-
3.	$1.00 \times 10^4 - 1.00 \times 10^7$	10	1b,2a,3a,4a	-	3	1	-	8	-	3	-
4.	$> 1.00 \times 10^8$	0	-	-	-	-	-	-	-	-	-
Total Cases =		24									

Table 2: Age wise HCV viral load and genotype distribution pattern

Age group in years	No. of Cases	HCV genotypes
0-20	1	3a
21-40	8	1b, 2a, 3a, 4a
41-60	14	1b, 3a, 4a
Above 60	1	Genotype not detected due to low viral load

Table 3: Percentage wise distribution of HCV genotypes

HCV genotype detected	No. of cases	Percentage
1a	-	-
1b	3	27.27%
2a	1	9.09%
2b	-	-
3a	8	72.72%
3b	-	-
4a	3	27.27%

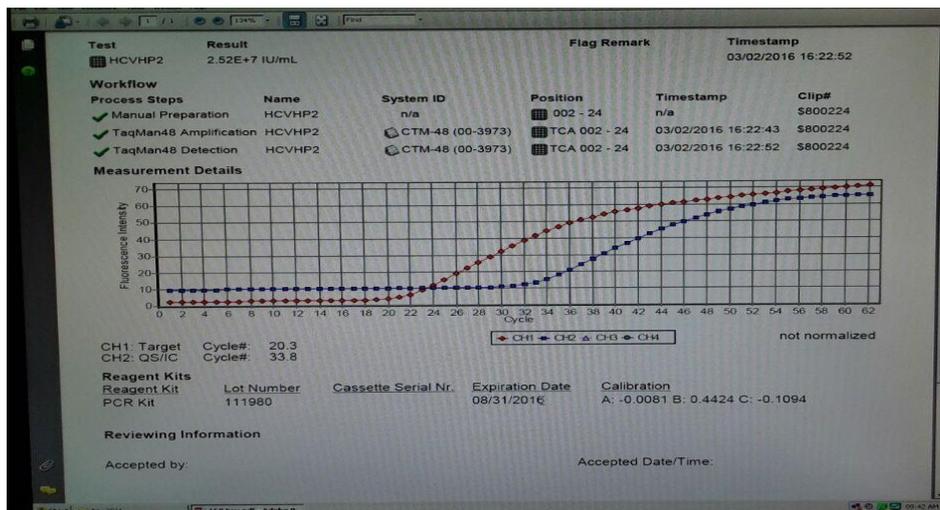


Fig 2: Amplification plot for HCV RNA viral load (Case= High HCV viral titer).

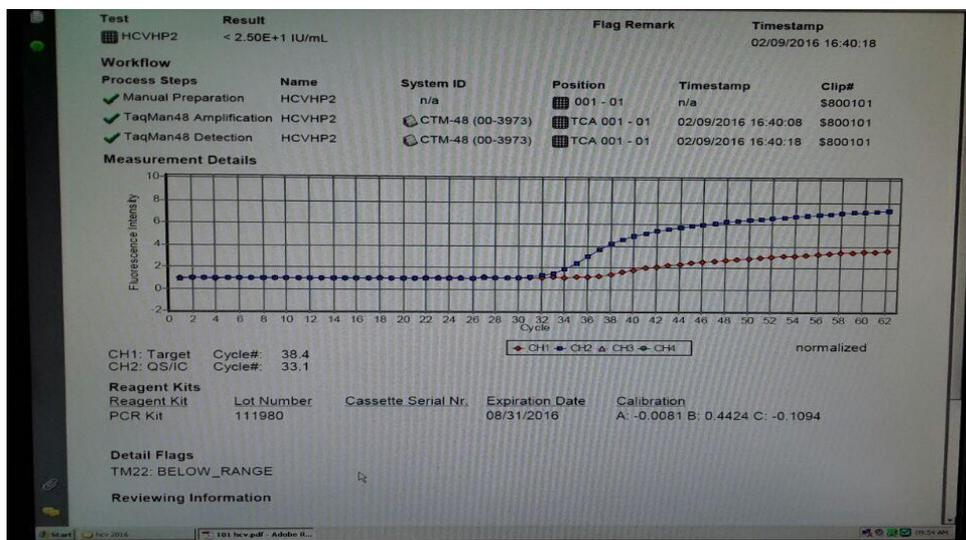


Fig 3: Amplification plot for HCV RNA viral load (Case= Low HCV viral titer)

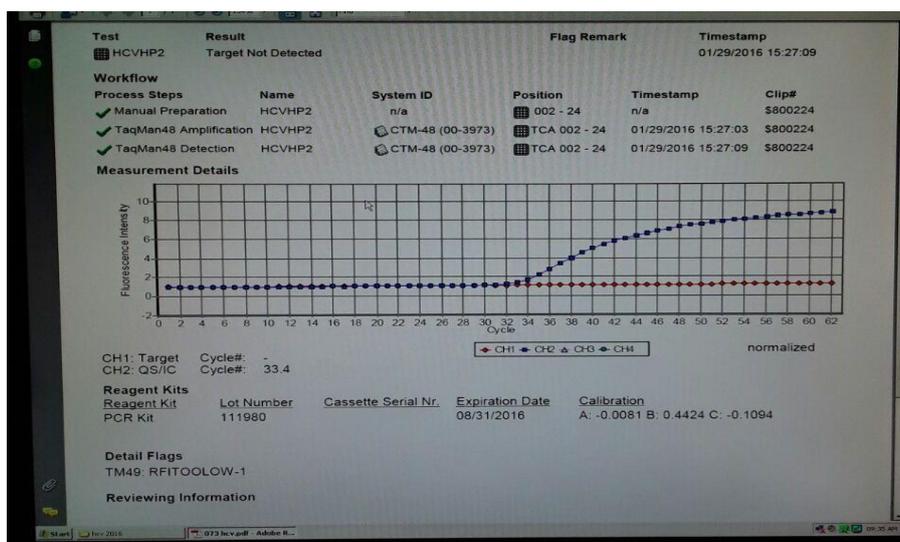


Fig 4: Amplification plot for HCV RNA viral load (Case= Target not detected)

DISCUSSION:

Differences among HCV genotypes in geographic distributions have provided investigators with an epidemiological marker that can be used to find the source of HCV infection in a given population & for further prognosis [20, 21]. The current study includes the collection of 24 specimens which were confirmed positive for HCV by serological findings. As the hospital includes the involvement of patients from hilly areas of Uttarakhand & Himachal Pradesh along with some parts of West U.P. Out of 11 samples which was processed for genotyping, 8 cases were for HCV genotype 3a of which 5 were males & 3 females, followed by HCV genotype 1b (3 males) & 4a (3 males). The genotype 3a was observed in 8 (72.72%) patients. Genotype 1b was seen in 3 (27.27%) patients. Genotype 4a was seen in 3 (27.27%) patients. Genotype 2a was seen in only 1 (9.09%) patient. Our findings include the prevalence of HCV genotype 3a in the clinical isolates, although HCV genotypes 1, 2 and 3 were also detected. From the current study genotype 3a was most prevalent and was present in males (the reason is not clear but it may be due to injection, drug abuse, needle stick accidents and transfusion of blood, intravenous drug users & tattooing etc). Similar study was also done by Pushpa Latha Manjunatha *et al.*; in 2015 and Chakravarti, Anita, Gaurav Dogra *et al.*; in 2011 [22-25]. This work can be further evaluated to study the exact HCV genotyping characterization in the Northern Indian population with more number of samples because this study was limited to the 24 samples only.

CONCLUSION:

The outcome of HCV genotyping is of almost clinical value as there are various regimens available to treat different types of HCV genotypes like Simeprevir,

Sofosbuvir etc for genotype 1. Sofosbuvir/R for genotype 2 & Sofosbuvir/R for genotype 3. There are various alternative therapies also available to treat Hepatitis C infection like Milk Thistle, Green tea extract, Glycyrrhizin but currently there is no vaccine available to prevent the Hepatitis C infection. The distribution of HCV genotypes vary according to the geographical region (26-28). The HCV genotype 3a is one of the most replicating viruses known to damage hepatic cells & thus requires proper line of treatment thoroughly during the diagnosis. Further the study can be done with more number of clinical isolates to get the exact epidemiological profiling of HCV genotypes.

Conflict of Interest: None

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REFERENCES:

1. Abdelwahab KS, Said ZN. Status of hepatitis C virus vaccination: Recent update. World journal of gastroenterology. 2016 Jan 14; 22(2):862.
2. Bailey C, Hern GH, Ortega RL, Sacchetti A, Strayer R. Hepatic Failure. *Emergency Medicine Practice+ Em Practice Guidelines Update. 2010 Apr 1; 12(4):1-22.*
3. Beaulieu PL, Tsantrizos YS. Inhibitors of the HCV NS5B polymerase: new hope for the treatment of hepatitis C infections. Current opinion in investigational drugs (London, England: 2000). 2004 Aug; 5(8):838-50.
4. Berry KE, Waghay S, Mortimer SA, Bai Y, Doudna JA. Crystal structure of the HCV IRES central domain reveals strategy for start-codon

- positioning. *Structure*. 2011 Oct 12; 19(10):1456-66.
5. Bustin SA. Absolute quantification of mRNA using real-time reverse transcription polymerase chain reaction assays. *Journal of molecular endocrinology*. 2000 Oct 1; 25(2):169-93.
 6. Bustin S. *AZ of Quantitative PCR*. (La Jolla, CA: International University Line).
 7. Carman WF, Hadziyannis S, McGarvey MJ, Jacyna MR, Karayiannis P, Makris A, Thomas HC. Mutation preventing formation of hepatitis B e antigen in patients with chronic hepatitis B infection. *The Lancet*. 1989 Sep 9; 334(8663):588-91.
 8. De Francesco R, Tomei L, Altamura S, Summa V, Migliaccio G. Approaching a new era for hepatitis C virus therapy: inhibitors of the NS3-4A serine protease and the NS5B RNA-dependent RNA polymerase. *Antiviral research*. 2003 Mar 31; 58(1):1-6.
 9. Didenko VV. DNA probes using fluorescence resonance energy transfer (FRET): designs and applications. *Bio techniques*. 2001 Nov; 31(5):1106.
 10. Dubuisson J. Hepatitis C virus proteins. *World Journal of Gastroenterology*. 2007 May 7; 13(17):2406.
 11. Forton DM, Allsop JM, Cox IJ, Hamilton G, Wesnes K, Thomas HC, Taylor-Robinson SD. A review of cognitive impairment and cerebral metabolite abnormalities in patients with hepatitis C infection. *Aids*. 2005 Oct 1; 19:S53-63.
 12. Fujisawa K, Tandon BN. Therapeutic approach to the chronic active liver disease: Summary of a satellite symposium. In *Viral hepatitis and liver disease 1994* (pp. 662-665). Springer Japan.
 13. Holland PM, Abramson RD, Watson R, Gelfand DH. Detection of specific polymerase chain reaction product by utilizing the 5' to 3' exonuclease activity of *Thermos aquaticus* DNA polymerase. *Proceedings of the National Academy of Sciences*. 1991 Aug 15; 88(16):7276-80.
 14. Idrees M, Riazuddin S. Frequency distribution of hepatitis C virus genotypes in different geographical regions of Pakistan and their possible routes of transmission. *BMC infectious diseases*. 2008 May 23; 8(1):1.
 15. Jubin R. Hepatitis C IRES: translating translation into a therapeutic target. *Current opinion in molecular therapeutics*. 2001 Jun; 3(3):278-87.
 16. Ronsin C, Pillet A, Bali C, Denoyel GA. Evaluation of the COBAS AmpliPrep-total nucleic acid isolation-COBAS TaqMan hepatitis B virus (HBV) quantitative test and comparison to the VERSANT HBV DNA 3.0 assay. *Journal of clinical microbiology*. 2006 Apr 1; 44(4):1390-9.
 17. Sarrazin C, Gärtner BC, Sizmman D, Babel R, Mihm U, Hofmann WP, von Wagner M, Zeuzem S. Comparison of conventional PCR with real-time PCR and branched DNA-based assays for hepatitis C virus RNA quantification and clinical significance for genotypes 1 to 5. *Journal of clinical microbiology*. 2006 Mar 1; 44(3):729-37.
 18. Widell A, Shev S, Månsson S, Zhang YY, Foberg U, Norkrans G, Fryden A, Weiland O, Kurkus J, Nordenfelt E. Genotyping of hepatitis C virus isolates by a modified polymerase chain reaction assay using type specific primers: epidemiological applications. *Journal of medical virology*. 1994 Nov 1; 44(3):272-9.
 19. Margolis HS, Coleman PJ, Brown RE, Mast EE, Sheingold SH, Arevalo JA. Prevention of hepatitis B virus transmission by immunization: an economic analysis of current recommendations. *Jama*. 1995 Oct 18; 274(15):1201-8.
 20. Available from: <http://www.mayoclinic.com/health/liver-failure/DS00961>. Retrieved May 20, 2011
 21. McOmish F, Yap PL, Dow BC, Follett EA, Seed C, Keller AJ, Cobain TJ, Krusius T, Kolho E, Naukkarinen R. Geographical distribution of hepatitis C virus genotypes in blood donors: an international collaborative survey. *Journal of Clinical Microbiology*. 1994 Apr 1; 32(4):884-92.
 22. Pushpalatha Manjunatha, Satish Kumar Amarnath, P. K. Menon, Arun Kumar H. R., Bala Satish M., S. J. Sabarish Babu. Molecular Epidemiology of Hepatitis C Virus and Predominant Genotype in India. *Clinical Medicine Research*. 2015; 4(5):139-142.
 23. Chakravarti A, Dogra G, Verma V, Srivastava AP. Distribution pattern of HCV genotypes & its association with viral load. *The Indian journal of medical research*. 2011 Mar 1; 133(3):326.
 24. Morishima C, Shuhart MC, Wang CC, Paschal DM, Apodaca MC, Liu Y, Sloan DD, Graf TN, Oberlies NH, Lee DY, Jerome KR. Silymarin inhibits in vitro T-cell proliferation and cytokine production in hepatitis C virus infection. *Gastroenterology*. 2010 Feb 28; 138(2):671-81.
 25. Nelson PK, Mathers BM, Cowie B, Hagan H, Des Jarlais D, Horyniak D, Degenhardt L. Global epidemiology of hepatitis B and hepatitis C in people who inject drugs: results of systematic reviews. *The Lancet*. 2011 Aug 19; 378(9791):571-83.
 26. Nousbaum JB, Pol S, Nalpas B, Landais P, Berthelot P, Brechot C. Hepatitis C virus type 1b (II) infection in France and Italy. *Annals of internal medicine*. 1995 Feb 1; 122(3):161-8.

27. Polyak SJ, Morishima C, Shuhart MC, Wang CC, Liu Y, Lee DY. Inhibition of T-cell inflammatory cytokines, hepatocyte NF- κ B signaling, and HCV infection by standardized silymarin. *Gastroenterology*. 2007 May 31; 132(5):1925-36.
28. Qi X, Su C, Ren W, Yang M, Jia J, Dai J, Xu W, Guo X. Association between portal vein thrombosis and risk of bleeding in liver cirrhosis: A systematic review of the literature. *Clinics and research in hepatology and gastroenterology*. 2015 Dec 31; 39(6):683-91.