# **Scholars Journal of Applied Medical Sciences (SJAMS)**

Abbreviated Key Title: Sch. J. App. Med. Sci. ©Scholars Academic and Scientific Publisher A Unit of Scholars Academic and Scientific Society, India www.saspublishers.com ISSN 2320-6691 (Online) ISSN 2347-954X (Print)

Biochemistry

# Beneficial Role of Ascorbic Acid against Lead Toxicity - A Mini Review

Ibrahim Abdulwaliyu<sup>1\*</sup>, Shefiat Olayemi Arekemase<sup>2</sup>, Musa Latayo Batari1 , Amina Ibrahim Madugu<sup>2</sup>, Olanipekun Oladele Idowu<sup>1</sup>, Aliyu Muhammad<sup>3</sup>, Oladipo Sunday Aribido<sup>1</sup>, Oluwayinka Olufunmilayo Owolabi<sup>4</sup>, Yakubu Sani<sup>5</sup>, Musa Halidu Mahmud<sup>1</sup>, and Suleiman Abdullahi<sup>6</sup>

<sup>1</sup>Department of Scientific and Industrial Research, National Research Institute for Chemical Technology, Basawa, Zaria, Nigeria

<sup>2</sup>Department of Environmental and Industrial Technology, National Research Institute for Chemical Technology, Basawa, Zaria, Nigeria

<sup>3</sup>Department of Biochemistry, Ahmadu Bello University, Zaria, Nigeria

<sup>4</sup>Bioresources Development Centre, National Biotechnology Development Agency, Lugbe, Abuja, Nigeria

<sup>5</sup>Department of Chemistry, Ahmadu Bello University, Zaria, Nigeria

<sup>6</sup>Directorate of Research and Development, Nigerian Institute of Science and Leather Technology, Zaria, Nigeria

## **Review Article**

\*Corresponding author Ibrahim Abdulwaliyu

**Article History** *Received:* 10.07.2018 *Accepted:* 15.07.2018 *Published:* 30.07.2018

**DOI:** 10.36347/sjams.2018.v06i07.021



**Abstract:** Environmental lead pollution is a continuous public health concern in Nigeria and the world at large. Exposure to lead results in toxicological complications, which range from brain disorders, decreased organ functions, and increased risk of chronic diseases especially in the elderly. Oxidative stress is the chief pathophysiologic mechanism by which lead impairs health integrity in humans and animals. So far, amongst the exogenous antioxidants, ascorbic acid is perhaps the most comprehensively studied antioxidant with respect to lead intoxication. It is capable of donating reducing equivalent to quench free radicals, and also facilitates lead excretion from the body. However, the bulk of the research on the role of ascorbic acid on lead toxicity was based on pre-treatment using animal models, while post treatment appears to be neglected. More so, despite enormous information on the role of ascorbic acid on lead toxicity, information on human clinical trials is still relatively scarce. This review provides a summary of toxicities associated with lead, and some benefits of ascorbic acid supplementation with respect to lead intoxication. **Keywords:** Vitamin C, Lead, Toxicity, Antioxidants, Oxidative stress.

#### Lead

Lead is a soft, grey-blue heavy metal found ubiquitously in nature, though at low concentration. It is widely dispersed into the environment, particularly as a result of human activities, an event that poses very serious clinical issues [1]. Such activities are common in areas where metals are mined, processed and used industrially [2].

Lead has been mined and smelted for at least 8000 years [3], and its toxic attributes have been recognized for more than 2000 years [4]. To date, lead still remains a critical environmental toxicant, despite concerted effort at tackling lead pollution.

Once introduced into the environment, it accumulates and persists [5], a scenario synonymously related to its pathophysiologic effects. Its persistent and non-biodegradable nature poses serious threats to human life [6], irrespective of age and gender. Sources of lead exposure include water, soil, dust, paints, batteries, leaded gasoline, food etc. [7, 8].

### ABBREVIATIONS

ALA: aminolevulinate; ALAD: aminolevulinate dehydratase; ASC: ascorbic acid; CaNa<sub>2</sub>EDTA: calcium disodium ethylenediamine tetra acetic acid; CAT: catalase; CNS: central nervous system; DMSA: dimercaptosuccinic acid; FSH: follicle stimulating hormone; GIT: gastrointestinal tract; GSH: reduced glutathione; GSHpx: glutathione peroxidase; GSSG: oxidized glutathione; H<sub>2</sub>O<sub>2</sub>: hydrogen peroxide; L: fatty acid radical; LDL: low density lipoprotein cholesterol; LH: fatty acid; LOO<sup>•</sup>: peroxyl radical; LOOH: hydroperoxide; NADP: nicotinamide adenine dinucleotide phosphate; NADPH: reduced nicotinamide adenine dinucleotide phosphate; MPO: myeloperoxidase; O<sup>\*</sup><sub>2</sub>: superoxide anion; OX-LDL: oxidized low density lipoprotein cholesterol; Pb: lead; PBG: porphobilinogen; PUFA: polyunsaturated

fatty acid; RDA: recommended dietary allowance; ROS: reactive oxygen species; SH: sulfhydryl; SOD: superoxide dismutase; SVCT<sub>1</sub>: sodium dependent vitamin C transport type 1; TC: total cholesterol; USA: United States of America; VLDLC: very low density lipoprotein cholesterol.

#### Ascorbic acid

Ascorbic acid is traditionally known as vitamin C, and structurally related to glucose [9]. It is well absorbed from the gastrointestinal tract (GIT), and the absorption takes place mainly in the distal intestine via ascorbate transportation, known as the sodium-dependent vitamin C transport type 1 (SVCT<sub>1</sub>). The absorbed ascorbic acid attains a plasma level of 50-60µm for a well-nourished, healthy non-smoker [10]. The recommended dietary allowance (RDA) of ascorbic acid for women and children is about 90mg/day and 45mg/day respectively [11]. Reduced intake of this vitamin is associated with scurvy development, anaemia, muscle degeneration, atherosclerotic plaques and infectious diseases [12]. Ascorbic acid is widely distributed in nature, especially in green leafy vegetables and fruits [13-15]. One of its biochemical roles is the modulation of collagen synthesis (it serves as a co-factor in the hydroxylation of proline and lysine residues) [16], and this makes it essential for wound healing [17, 18]. It is also important for carnithine and neurotransmitter biosynthesis [19]. Ascorbic acid also participates in metal catalysed reactions such that it reduces oxides of iron (Fe<sup>3+</sup>) and copper (Cu<sup>2+</sup>) respectively [20]. It helps to regenerate some antioxidants such as Vitamin E and β-carotene [21-24] both of which possess anti-oxidative stress properties. Ascorbic acid has both industrial and medicinal values. It prevents loss of flavour and aroma, extends shell life and enhances nutrient content of processed foods [25]. Its medicinal attributes include anti-atherogenic, anti-cancer, anti-inflammatory properties; it helps (mega doses) in the prevention and treatment of cataracts, glaucoma, stroke, diabetes etc. [26-30].



Fig-1: Structure ascorbic acid. C<sub>6</sub>H<sub>8</sub>O<sub>6</sub>

The relationship between ascorbic acid and lead toxicity is beyond recent time. Ascorbic acid has the ability to modulate lead induced toxicity, though preventive measures have remained the best solution to curbing the menace associated with lead toxicity. But despite efforts made to tackling the threat, both occupational and environmental lead exposure remain a serious threat, particularly in developing nations, where uncontrolled mining activities still take place. More so, the multifaceted effects of lead on biochemical processes, and overall pathophysiologic ill nature of lead remain a big challenge. Therefore, this review focus on the role of ascorbic acid on toxicities associated with lead.

### Benefits of ascorbic acid on lead induced oxidative Stress

### Lead induced oxidative stress

Lead induced oxidative stress has been postulated as the basic mechanism by which lead exerts toxic effects [31-35]. Oxidative stress represents an imbalance between free radical production, resulting in various degree of body injury, and inability of the body's system to repair the damage. It is associated with drastic increase in the malonyldialdehyde (a measure of lipid peroxidation) content, and decrease in the activities of endogenous antioxidants such as superoxide dismutase (SOD), catalase (CAT), glutathione (GSH) etc. [36, 37].



Fig-2: Proposed mechanism of lead induced toxicity. In ideal physiological condition, free radicals generated during oxygen metabolic process exist as nontoxic substances as their levels are being regulated by endogenous enzymes, such as catalase (CAT), superoxide dismutase (SOD), and glutathione peroxidase (GSHpx). However, exposure to lead decreased the activities of the aforementioned antioxidants, thereby leading to generation of reactive oxygen species (ROS), molecules capable of inducing various degrees of body injuries.

Oxygen consumption is vital especially in aerobic metabolic process, where it is reduced to water. Lead amongst other factor interferes with the process via three mechanistic approach; (1) the displacement of antioxidants co-factors [38, 39], (2) inhibition of the heme synthesis pathway, (3) covalent attachment to sulfhydryl (-SH) containing molecules (Figure 1).

#### Lead and superoxide dismutase (SOD) activity

Under aerobic condition, oxygen is converted to superoxide by NADPH oxidase, using reduced nicotinamide adenine dinucleotide phosphate (NADPH) as substrate. The NADPH transfers electron to the molecular oxygen ( $O_2$ ) to produce superoxide anion ( $O_2$ ), a reactive free radical.

 $NADPH + 2O_2 \stackrel{Oxidase}{\longleftrightarrow} NADP^+ + 2O_2^- + H^+$ 



Fig-3: Proposed mechanism of lead induced oxidative stress, and role of ascorbic acid. CAT- catalase, GSHpxglutathione peroxidase, H<sub>2</sub>O<sub>2</sub>-hydrogen peroxide, NADP-nicotinamide adenine dinucleotide phosphate, NADPHreduced nicotinamide adenine dinucleotide phosphate, MPO-myeloperoxidase, O<sup>-</sup><sub>2</sub>-superoxide anion, Pb-lead, SOD-superoxide dismutase. H<sub>2</sub>O- water, HOCL- hypochlorous acid, ASC- ascorbic acid, (-) inhibition.

The O<sup>-2</sup> generated, is a primary byproduct of oxygen metabolism [40], which progresses to the formation of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) by SOD. This enzyme contains copper and zinc as co-factors. Both are important for the functionality and stability of SOD. However, the functionality and stability could be impaired in the presence of lead, thereby rendering the enzyme less active or inactive, so the conversion of O<sup>-2</sup> to H<sub>2</sub>O<sub>2</sub> is impeded.  $Cu^{2+} - SOD + O_2^- \rightarrow Cu^+ - SOD + O_2$  $Cu^+ - SOD + O_2^- + 2H^+ \rightarrow Cu^{2+} - SOD + H_2O_2$ 

#### Lead and catalase (CAT) activity

Lead has also shown to impair the activity of catalase (CAT), the enzyme that catalyses the conversion of  $H_2O_2$  to metabolic water (H<sub>2</sub>O) and molecular oxygen (O<sub>2</sub>). It impairs CAT function via inhibition of heme biosynthesis [39]. It inhibits  $\delta$ -aminolevulinate synthase, the enzyme that catalyses the condensation of glycine (a non-essential amino acid), and succinyl Co-A (intermediate of the TCA cycle) to  $\delta$ -aminolevulinate (ALA). Lead also inhibits the activities of  $\delta$ -aminolevulinate dehydratase (ALAD), an enzyme that condenses two molecules of  $\delta$ -aminolevulinate to porphobilinogen (PBG). It either binds to the sulfhydryl (-SH) component of the enzyme (ALAD), or displaces the enzyme co-factor (zinc) [41], thereby rendering the enzyme inactive. More so, lead inhibits the activities of ferrous chelatase (ferrous synthetase), the enzyme responsible for the insertion of ferrous iron (Fe<sup>2+</sup>) into the central cavity of protoporphyrin IX, a metabolic event that leads to the formation of heme. Heme groups are very important for the functionality of CAT.

 $\begin{array}{l} 2H_2O_2 \xrightarrow{catalase} 2H_2O + O_2 \\ H_2O_2 + Fe^{3+} - CAT \rightarrow H_2O + O = Fe^{4+} - CAT(\bullet +) \\ H_2O_2 + O = Fe^{4+} - CAT(\bullet +) \rightarrow H_2O + Fe^{3+} - CAT + O_2 \end{array}$ 

The reaction takes place via the interaction of  $H_2O_2$  with asparagine and histidine of the amino acids sequence of CAT, and allows the transfer of a proton (hydrogen ion) between the oxygen atoms. The oxygen atom co-ordinates with the iron, and frees the water molecule. The heme containing enzyme then reacts with another  $H_2O_2$  molecules to reform the Fe<sup>3+</sup>-CAT,  $H_2O$  and  $O_2$  respectively. It's worthy to note that the Fe<sup>3+</sup> represent the iron containing heme, attached to the enzyme. The Fe<sup>4+</sup> formed is not relatively stable, so it receives stabilizing electron (a radical cation ( $\bullet$ +) to stabilize itself. So, the overall effect of lead on heme synthesis consequently impairs the conversion of  $H_2O_2$  to  $H_2O$  and  $O_2$ . This implies that there will be more of  $H_2O_2$  generation than it decomposition.

### Lead and glutathione peroxidase activity

Glutathione peroxidase (GSHpx), another endogenous enzyme has the ability to convert  $H_2O_2$  to  $H_2O$ , using glutathione (GSH) as substrate. However, exposure to lead could impede the activities of GSHpx via the displacement of

selenium, a key co-factor of the enzyme, thereby making the enzyme inactive. More so, it covalently attaches itself to the sulfhydryl groups of the substrate, which eventually get degenerated [39].

 $2G - SH + H_2O_2 \rightarrow GS - SG + 2H_2O$ Thus, the metabolic arrest of GSHpx and CAT by lead hampers the decomposition of H<sub>2</sub>O<sub>2</sub> to H<sub>2</sub>O, and the H<sub>2</sub>O<sub>2</sub> instead progresses to hydroxyl radical (•OH) formation.

 $\begin{array}{rcl} Fe^{2+} + H_2O_2 & \rightarrow & Fe^{3+} + HO^{\bullet} + OH^- \\ Fe^{3+} + H_2O_2 & \rightarrow & Fe^{2+} + HOO^{\bullet} + H^+ \end{array}$ 

The hydroxyl radical can initiate a sequence of reactions that could lead to membrane degradation.  $LH + HO^{\bullet} \rightarrow L^{\bullet} + H_2O$ 

The  $^{\bullet}$ OH is chemically unstable. It removes hydrogen atom (H) from the vulnerable site (double bond) of polyunsaturated fatty acids (PUFA), represented as LH, thereby producing fatty acid radical (L $^{\bullet}$ ). Under aerobic conditions, the L $^{\bullet}$  takes up oxygen to produce peroxyl radical (LOO $^{\bullet}$ ). The LOO $^{\bullet}$  can also attract hydrogen atom to form hydroperoxide (LOOH), and these reactions continues in series, leading to oxidation of PUFA.

 $\begin{array}{rcl} L^{\bullet} + O_2 & \rightarrow & LOO^{\bullet} \\ LOO^{\bullet} + LH & \rightarrow & LOOH + L^{\bullet} \end{array}$ 

### Role of ascorbic acid on oxidative stress

Ascorbic acid has the ability to normalize alteration of oxidative stress biomarkers initiated by lead [42-45]. Its ability to quench free radicals and chelate heavy metals makes it a unique antioxidant [46], and this led to the presumption that supplementation of ascorbic acid could be the best chelation therapy for lead intoxication [47]. This was further strengthened in studies by Wang *et al.* [37], and Seven *et al.* [48] that lead intoxication was shown to reduce the level of endogenous antioxidants which was normalized upon ascorbic acid supplementation. Ascorbic acid does not only reduce or reverse oxidative stress, but also helps to replenish and improve ascorbic acid level [49], which is vital for maximum health integrity.

Depletion of antioxidants by lead leads to proliferation of reactive oxygen species (ROS) ( $O_2$ ,  $H_2O_2$ ,  $\bullet OH$ ) [50], most of which could be scavenged by ascorbic acid supplementation [12] directly or indirectly. Ascorbic acid directly scavenges free radicals, either generated by lead or by other factors [51, 52], and protects the cells from oxidative damage [53, 54]. The hydrogen atoms of ascorbic acid pairs up with unpaired electron of the free radicals, converting them to non-free radicals.

 $2O_2^- + 2H + ascorbate \rightarrow 2H_2O_2 + dehdroascorbate$  $H_2O_2 + 2ascorbate \rightarrow 2H_2O + 2monodehydroascorbate$ 

Ascorbic acid indirectly quenches free radicals by virtue of regenerating some important antioxidants such as GSH, and vitamin E [52]. The latter is capable of terminating lipid peroxidation chain reactions [55].

 $\begin{array}{rcl} ASCH_2 + GS - SG & \rightarrow & ASC + 2GSH \\ LOO^{\bullet} + \alpha - to copherol - OH & \rightarrow & LOOH + \alpha - to copherol - O \end{array}$ 

Name		
Singlet oxygen		
Super anion radical		
Hydroxyl radical		
Alkoxyl radical		
Peroxyl radical		
Hydrogen peroxide		
Hydroperoxide		

### Table-1: List of ROS [55].

#### **Consequences of oxidative stress**

Under normal physiological conditions, the aforementioned reactive species are not harmful to the body. However, when the production of ROS exceeds the cellular antioxidant capacity, it becomes harmful to the host. ROS destroys biomolecules (lipids, proteins, nucleic acids), and predisposes the host to health complications such as neurodegenerative diseases (Alzheimer's disease, Parkinson's disease etc.) [56-59], cancer [60-62], and diabetes [63-65].

#### Ascorbic acid and cancer

The antioxidant role of ascorbic acid is vital to processes associated with cancer [66], although the relationship between ascorbic acid and cancer development are controversially related. Late stage cancer patients usually have ascorbic acid deficiencies [67], hence the need from exogenous source becomes inevitable, as it plays a vital role in improving quality of health [68]. Controversially, Gonzalez *et al.* [16] explicitly stated that many scientists failed to reproduce the scientific basis that was earlier presented that ascorbic acid can be used as a therapeutic agent in the treatment of cancer.

Lead	Children	Adults
toxicity		
Acute toxicity	Headache [69], abdominal pain, vomiting, muscle pain, irritability, attention deficit, constipation, seizure etc.	High blood pressure, headache [69], vomiting, muscle pain, abdominal pain [70].
Chronic toxicity	Anemia (haemolytic anemia and frank anemia) [39], hearing loss, development delay, stunted growth.	Neuronal problem, reduced sperm count, repeated miscarriage [71], still birth, diabetes, cancer, stroke, arthritis, Organs (kidney, brain etc.) damage [72].

Table-2: Some symptoms and health issues associated with lead exposure

### Ascorbic acid and diabetes

The displacement of some essential metals by lead, could in part explain the pathophysiologic mechanism by which lead induces diabetes or processes associated with diabetes. Most essential metals, though not produced by the body, are necessary for maximum health integrity. For example, chromium ( $Cr^{3+}$ ), which performs insulin like functions [73], and zinc are important for the optimal activity of insulin, in terms of secretion and regulation of blood glucose levels. Since lead exhibits the ability to displace the aforementioned essential metals, then insulin secretion and function could be impaired. More so, the onset of oxidative stress may decrease the activity of insulin gene promoter and mRNA expression in pancreatic islet cells [74]. The impairment of insulin secretion and function shoots blood glucose level, which may probably exceed normal range of 126mg/dl, and persistent increase could lead to chronic hyperglycaemia [75]. The onset of hyperglycaemia exacerbates ascorbic acid deficiency, as it competitively inhibits glucose transport system [76], also responsible for the transport of oxidized form of ascorbic acid to cells. This inhibition may contribute to oxidative stress and consequently increase risk of cardiovascular diseases [76].

#### Ascorbic acid and cardiovascular disease risk

Oxidation of low density lipoprotein cholesterol (OX-LDLC), may partly explain oxidative stress induced cardiovascular diseases [77-81]. OX-LDLC, being that it is a target molecule for scavenging receptors, could easily be incorporated in to plaque formation [82]. A scenario that contributes to narrowing of arterial blood vessels, thereby contributing to cardiovascular health risk. However, ascorbic acid has shown to prevent the oxidation of LDLC, and decrease the risk of cardiovascular health issues [83].

### Advantages of ascorbic over conventional chelating therapy

Ascorbic acid is probably the most extensively studied antioxidant with respect to lead induced oxidative stress [39], metabolic event proposed to be the main factor responsible for organ damage [36]. It has advantages over conventional chelating agents (DMSA, CaNa<sub>2</sub>EDTA). The conventional chelating agents are mostly used in the removal of lead from the body, especially in acute lead exposure, but ineffective in reversing metabolic injuries associated with lead induced oxidative stress. They have deleterious side effects [47], including disrupting essential micronutrient balance and enhancing the redistribution of lead from the stored site (bone) to other regions (brain, liver, testes, kidney etc.) of the body [36]. Such redistribution may further aggravate organ dysfunction, especially in pre-existing cases of organ malfunction. Moreso, conventional chelating agents may also not be useful in instances of chronic lead exposure, where people are exposed to the toxic metal over a long period of time. Such exposure (chronic) usually manifests irreversible health complications, which could be managed by ascorbic acid supplementation, as it could easily be excreted from the body, and poses little or no health risk.

However, continuous use of mega doses of ascorbic acid may put the kidney at risk. It contributes to oxalic acid formation, the end product of ascorbic acid oxidation that has the potential to crystallise as calcium oxalate in the urinary space [84] thereby increasing the risk of kidney stone formation [85, 86]. It may also serve as a pro-oxidant (in mega doses), and heighten oxidative stress rather than mitigating it. As a pro-oxidant, ascorbic acid interacts with transition metal ions and promotes their reduction, accompanied by increased  $H_2O_2$  production and consequently **°OH** formation [87].

#### Benefit of ascorbic acid on lead induced hepatotoxicity

The liver, being the primary metabolic organ of the body, is highly vulnerable to lead intoxication [88-92]. However, ascorbic acid supplementation has been shown to mitigate lead induced hepatic damage [3, 93-95], chiefly in animal models.

#### Benefit of ascorbic acid on lead induced infertility

The toxic insult of lead to the reproductive system is one of the fastest growing research interests in toxicological discipline. Maternal lead exposure during neonatal period has dose-related and long term effects on postnatal development of testes as seen in offspring of Wister rats [96], and this maybe the possible cause of infertility seen in adulthood. Sometimes male infertility of unknown aetiology may be attributed to various environmental and occupational exposures to toxic substances, such as lead [97]. Other complications associated with maternal lead exposure include decreased birth weight, morphological abnormalities in the head and limbs, [38, 98, 99], spontaneous abortions, stillbirth and miscarriage [99, 100]. Exposure to lead could also cause reduction in the serum levels of gonadotropins such as the follicle stimulating hormone (FSH), luteinizing hormone (LH), testosterone [101], reduction of spermatogenic cells, and reduction of the size and diameter of seminiferous tubules [102, 103], all of which are vital for the maintenance of testicular activity.

Lead induced testicular damage is a function of ROS generation [104-106], that has been linked to decreased sperm integrity [107], germ cell death, low sperm synthesis (hypospermatogenesis) and, above all, testicular damage [108]. However, ascorbic acid is an antioxidant with long known fertility importance. It has been shown to improve sperm quality [109], human fertility [110], increase spermatogenesis and maintain the volume of testes of lead intoxicated Wister rats [111]. A study by [112] showed that lead treated mice exhibit deformations of sperm morphology and testicular injury, while daily supplementation of ascorbic acid improves the sperm morphology. More so, ascorbic acid supplementation has been shown to modulate toxicity associated with prolonged lead exposure [113]. Similarly, Ayinde *et al.* [31] verified the influence of ascorbic acid on testicular zinc content and testicular damage in lead exposed albino rats. Their findings revealed that lead intoxication was responsible for histological damage and disturbances of male reproductive organs. However, ameliorative effects were observed upon vitamin C and/or vitamin E supplementation. Supplemented ascorbic acid concentrates appreciably in the seminal plasma of living species [114], where it protects the testes [115], and maintains the genetic architecture of the sperm cells [116]. Thus, decrease in the concentration of testicular ascorbic acid content may predispose the testes to toxic injury.

Despite the described, significant role of ascorbic acid on the maintenance of the reproductive integrity, excessive intake has been linked to reproductive failure, while deficiency decline reproductive performance in Wister rats [114]. This implies that both deficiency and excessive intake of ascorbic acid affect reproductive performance. However, low dosage of ascorbic acid, co-supplemented with other vitamins has been shown to be more effectual. Wang *et al.* [117] explored the impact of combined administration of ascorbic acid and thiamine (at different levels) on the apoptosis of lead exposed mice testes. The impaired testicular tissues were ameliorated by the lower doses of ascorbic acid and thiamine, while the highest dose of the co-supplementation promotes testicular cell death.

#### Benefit of ascorbic acid on lead induced dyslipidaemia

Exposure to lead has been shown to induce lipid abnormalities, and increase risk of atherosclerosis [118-121]. Atherosclerosis is a condition in which plaque (made up of fat, cholesterol, calcium and other substance found in the blood) builds up inside the arteries (blood vessels that carries oxygen rich blood to heart and away from the heart), thereby impeding blood flow. Any obstruction in the blood flow could lead to health problems such as heart attack, stroke, or even death. Lead induced hyperlipidaemia is a potential, though modifiable risk factor for coronary artery disease, the leading cause of death in developed countries (e.g. USA) [122-124]. However, curbing such effects has been promising using various chelating agents such as ascorbic acid. Ugbaja *et al.* [125] investigated the comparative effect of ascorbic acid and conventional chelation therapy on lead induced dyslipidaemia. They revealed that ascorbic acid may not be more efficacious than other chelating agents but could be a cheaper and more convenient therapy for lead toxicity. It possesses the ability to lower total cholesterol (TC) level, very low density lipoprotein cholesterol (VLDLC) and low density lipoprotein cholesterol (LDLC) [126]. More so, ascorbic acid deficiency is an inevitable contributing factor for cardiovascular disease and of course a risk factor for cardiovascular related morbidity [127].

### Benefit of ascorbic acid on lead induced alterations of haematological indices

Exposure to lead, especially during the first few days, impairs the hematopoietic system [4, 128-130]. Lead acetate of environmentally comparable concentrations induced haematological changes, of which significant reduction in pack cell volume, haemoglobin concentration and a significant increase in total leukocyte counts in albino rats were noteworthy [2]. Among the haematological parameters, the erythrocytes exhibited a high affinity for lead, thus making them more vulnerable than other haematological indices [131]. The implication of lead on haematological architecture includes anaemia, and decreased immune working capacity. However, ascorbic acid could efficiently enhance the

working capacity of the immune system by a mechanism that involves the combination of humoral, immune competence, and cell mediated defence reaction [132]. The cell mediated immunity of the ascorbate is a function of increased ascorbate contents of the leukocytes [133].

### Benefit of ascorbic acid on lead induced brain damage

The brain exhibits high degree of vulnerability to lead intoxication, especially in children with neurological challenges [134]. The chief target of lead toxicity is the central nervous system (CNS), where it causes permanent intellectual deficit, such as behavioural and learning abnormalities, cognitive impairment, memory loss etc. [135-139]. Exposure to lead especially during pregnancy and lactation could be responsible for behavioural and cognitive impairment in infants [140]. However, ascorbic acid has shown to be an important antioxidant in the protection of the brain cells, especially in oxidative brain damage [141, 142]. Salehi *et al.* [143] investigated the detrimental role of lead on learning and memory loss, and the possible preventive role of ascorbic acid. Their findings revealed that lead treatment impairs learning and memory, while ascorbic supplementation improves learning and reduces memory deficit pre and post exposure to lead. More so, Musa *et al.* [144] examined the protective role of ascorbic acid on the cerebellum of lead intoxicated adult Wister rats. Their findings clearly revealed that lead exposure causes significant cerebellum degeneration in the brain of the experimental rats, and supplementation of ascorbic acid counteracts the damage.

### CONCLUSION

Lead is a serious environmental toxicant that affects almost every living creature. The role of vitamin C in lead intoxication is well documented across the globe. However, research so far has primarily focused on the protective and ameliorative effect of ascorbic acid on lead intoxication. More so, studies on human clinical trials are relatively scarce. In light of these, we suggest that increased focus should be placed on explorations of other areas, including tandem action of ascorbic acid with other chelating agents or effects of ascorbic acid on sequestered heavy metals. These may open up new avenues for the treatment of chronically lead intoxicated patients.

### REFERENCES

- 1. Pokras MA, Kueeland MR. Lead poisoning: using trans disciplinary approaches to Solve ancient problems. Eco Health 2008; 5(3):379-385.
- 2. Ekanem AU, Kwari HD, Garba SH, Salami HA. Effect of lead acetate on spleen and blood parameters in albino Rats. J Dental Med Sci. 2015; 14(3):43-49.
- 3. Suleiman JB, Eze ED, Momoh IJ, Usman W, Hedima NC, Zipele HM et al. Ameliorative effect of vitamin C on serum liver enzymes in lead induced toxicity in Wister Rats. J Sci 2013; 3(1): 188-192.
- 4. Mannem P. Lead Toxicity on haematological changes and amelioration with Ginger (*Zingiber officianale*) extract in male albino rats. Internat. J. Adv. Res 2014; 2(4):23-28.
- 5. Sainath SD, Meena R, Supriya CH, Reddy KP, Reddy PS. Protective role of *Centella asiatica* on lead induced oxidative stress and suppressed reproductive health in male rats. Env Toxicol Pharmacol. 2011; 32:146-154.
- 6. Vig EK, Hu H. Lead toxicity in older adults. J Am Geriatr Sci. 2000; 48 (11): 1501-6.
- 7. Chowdhury RA. Recent advances in heavy metals induced effects on male reproductive function A retrospective. Al Ameen J Med Sci. 2009; 2(2): 37-42.
- 8. Garaza A, Vega R, Soto E. Cellular mechanism of lead neurotoxicity. Med Sci Monitor 2006; 12 (3):57-65.
- 9. Adikwu E, Deo. Hepatoprotective effect of vitamin C. J Pharmacol Pharm. 2013; 4:84-92.
- 10. Duarte TL, Lunec J. When an antioxidant is not an antioxidant? A review of novel actions and reactions of vitamin C. J Free Radic Res. 2005; 39(7):671-86.
- 11. Philip KM, Terrago-Trani MT, Gebhardi SE, Exler J, Putterson KY, Haytowitz DB et al. Stability of vitamin C in frozen raw fruits and vegetable homogenates. J Fd Com Analy. 2010; 23: 253-259.
- 12. Subasree S. Role of vitamin C and vitamin E in health and disease. J Pharmaceut Sci Res. 2014; 6 (1): 52-55.
- 13. Chatterjea MN, Shinde R. Textbook of Medicinal Biochemistry 5<sup>th</sup> edition. AYPE 2002; 154-157.
- 14. Mathiventhan U, Ramiah S. Vitamin C content of commonly eaten green leafy vegetables in fresh and under different storage conditions. Tropical plant Res.2015; 2(3): 240-245.
- 15. Pacier C, Martirosyan DM. Vitamin C: Optimal Dosages, Supplementation and use in Disease Prevention. Funct Fds Health Dis. 2015; 5(3):89-107.
- 16. Gonzalez MJ, Massari JRM, Mora EM, Guzman A, Riodan NH, Riodan HD et al. Orthomolecular Oncology Review: Ascorbic acid and Cancer 25 years later. Integ Cancer Therap. 2005; 4(1): 32 44.
- 17. Kamer E, Unalp HR, Gundogan O, Diniz G, Ortac R, Olukman M et al. Effect of ascorbic acid on incisional wound healing in streptozotocin induced diabetic rats. J Wounds. 2010; 22 (2): 27-31.
- 18. Gross RL. The effect of ascorbate on wound healing. J Intopthalmol Clini 2000; 40 (4): 51-56.
- 19. Zeraati F, Araghchian M, Esna ashari F, Fazliais MM, Torabian S, Fallah N et al. Antinociceptive properties of ascorbic acid. Evidence of the mechanism of action. J Med Biochem. 2014; 2(1)8572.
- 20. Kuo SM. The multifaceted biological role of vitamin C. J Nutri Fd Sci. 2013; 3(5): 231-236.

- 21. Aburawi SM, Aghambirion F, Attom AA, Alfubuly RA, Kara AA. Effect of ascorbic acid on mental depression drug therapy. J Psycho Psychother. 2014; 4: 131-9.
- 22. Hacisevki A. An overview of ascorbic acid biochemistry. J Fac Pharm Ankara 2009; 38(3): 233-255.
- 23. Mazid M, Khan TA, Khan ZH, Qudusi S, Mohammed F. Occurrence, biosynthesis and potentialities of ascorbic acid in plants. Intern. *J Plant Ani Env Sci.* 2011; 1(2):167-185.
- 24. Skrovankova S, Micek J, Sochor J, Baron M, Kynicky J, Jurikova J. Determination of ascorbic acid by electrochemical techniques and other methods. Internat J Electr Sci. 2015; 10:2421-2431.
- 25. Hancock RD. Recent patents on vitamin C: Opportunity for crop improvement and single step biological malfunction. Recent patent food. Nutr Agric. 2009; 1: 39-49.
- 26. Evangelou A, Kalfakakou V, Georgakas P, Koutrass V, Vezyrati P, Illopoulou L. Effect on withdrawal syndrome of heroin abusers. Invivo J. 2000; 14 (3):363-6.
- 27. Iqbal K, Khan A, Khattack AKM. Biological significance of ascorbic acid (vitamin C) in human health- A reviewer. Afri J Fd Agric Nutr Dev. 2004; 5 (1): 1-14.
- 28. Naidu KA. Vitamin C in human Health and disease is still a mystery: an Overview. Nutri J. 2003; 2:7-15.
- 29. Padaytty SJ, Levine M. New insight into the Physiology and Pharmacology of vitamin C. CMAJ 2001; 164(3):353-4.
- 30. Walingo M. Role of ascorbic acid on human health A review. Afri J Fd Agric Nutr Dev 2005; 5(1): 1-4.
- 31. Ayinde O, Ogunnowo S, Ogedegbe RA. Influence of vitamin C and E on testicular zinc content and testicular toxicity in lead exposed albino rats. Pharmacol Toxicol. 2012; 13:17.
- 32. Moniem A, Dkhil M, Al-Quraishy S. Protective role of flaxseed Oil against lead induced oxidative stress in testes of adult rats. Afri J Biotech. 2010; 9(42):7216-7223.
- 33. Pande M, Mehta A, Pant BP, Flora SJS. Combined administration of chelating agent and an antioxidant in the prevention and treatment of acute lead intoxication in rats. J Env Toxicol Pharmacol. 2001; 9:173-184.
- 34. Sharma S, Sharma V, Paliwal R. Lead toxicity oxidative damage and health implcation: A Review. Internat J Biotech Molecul Biol Res. 2011; 2(13):215-221.
- 35. Xu J, Ling JL, Chen WU, Xio FW, Wenyu FU, Lihong X. Lead induces oxidative stress, DNA damage and alteration of P53, Bax and Bcl-2 Expression in Mice. Fd Chem Toxic 2008; 46:1488-1494.
- 36. Ebuehi OAT, Ogedegbe RA, Ebuehi OM. Oral administration of vitamin C and E ameliorates lead induced hepatotoxicity and oxidative stress in albino rats. Nige. Qt J. Hosp. Med 2012; 22(2):85-90.
- 37. Wang C, Liang J, Zhang C, Bi Y, Shi XM, Shi Q. Effect of ascorbic acid and thiamin supplementation at different concentration on lead toxicity in rats. Ann Occup Hyg. 2007; 51(6):563-569.
- 38. Aprioku JS, Siminialayi IM. Maternal lead exposure and pregnancy outcome in Wister rats. J Toxicol Env Health Sci 2013; 5(10):185-193.
- 39. Flora G, Gupta D, Tiwari A. Toxicity of lead: A review with recent updates. J Interdisci 2012; 5(2): 47-58.
- 40. Jalilov AS, Zhang C, Samuel ELG, Sikkeema WKA, Wu G, Berka V et al. Mechanistic study of the conversion of superoxide to oxygen and hydrogen peroxide in carbon nano partcles. Appl Mater Interfaces. 2016; 8:15086-15092.
- 41. Ercal N, Gurer OH, Aykin-Burns N. Toxic metals and oxidative stress: Mechanisms involved in metal induced oxidative stress. Curr Topics Med Chem. 2001; 1:529-539.
- 42. Chang BJ, Jang BJ, Son TG, Cho IH, Quan FS, Choe NH. Ascorbic acid ameliorates oxidative damage Induced by maternal low level lead exposure in the hippocampus of rat's pups during gestation and lactation. J Fd Chem Toxicol. 2011; 52:104-108.
- 43. Ferreira AG, Stefanello FM, Cunha AA, da Cunha MJ, Pereira TC, Bonan CD, Bogo MR, Netto CA, Wyse AT. Role of antioxidants on Na+, K+-ATPase activity and gene expression in cerebral cortex of hyperprolinemic rats. Metabolic brain disease. 2011 Jun 1;26(2):141-7.
- 44. Ghanwat G, Patil A, Patil J, Kshirsagar M, Sontakke A, Ayachit RK. Effects of vitamin C supplementation on blood level, oxidative stress and antioxidant status of battery manufacturing workers of western Maharashtra, India. J Clin Diag Res. 2016; 10(4):8-11.
- 45. Jewo PI, Duru FI, Fadeyibi IO, Saalu LC, Noronha CC. The protective role of ascorbic acid in burn induced testicular damage in rats. J Burns. 2012; 38(1):113-9.
- 46. Dass KK, Saha S. L-Ascorbic acid and alpha tocopherol supplementation and antioxidant status in nickel or lead exposed rat's brain tissue. J Basic Clin Physiol Pharmacol. 2010; 21:325-346.
- 47. Flora S, Pande M, Mehta A. Beneficial effect of combined administration of some naturally occurring antioxidants vitamins and their chelators in the treatment of chronic lead intoxication. Chem Bio Interact. 2003; 145: 267-280.
- 48. Seven I, Aksu T, Seven PT. The effect of Proposlis on biochemical parameters and activity of antioxidants enzymes in broilers exposed to lead induced oxidative stress. Asian Australian J Ani Sci. 2010; 23(11):1482-1489.
- 49. Ramdas BV. Protective role of ascorbic acid on the lead chloride Induced alterations in the ascorbic content of the fresh water fish, *Channa orientalis* (Schneider). Central. Eur J Exp Bio. 2013; 2(4): 34-37.

- 50. Ambali SF, Angali M, Shittu M, Kawu MU. Haematological changes induced by subchronic co-administration of chlorpyrifos and lead in Wister rats. Alleviating effect of vitamin C. Der Pharmacia Sinica. 2011; 2(2):276-284.
- 51. Washio K, Inagaki M, Tsuji M, Morio Y, Akiyama S, Gotoh H et al. Oral vitamin C supplementation in hemodialysis patients and its effects on the plasma level of oxidized ascorbic acid and Cu/Zn superoxide dismutase on oxidative stress markers. J Nephron Clin Pract. 2008; 109:49-54.
- 52. Jagetia GC, Rajanikant GK, Rao SK, Baliga MS. Alteration in the glutathione, glutathione peroxidase, superoxide dismutase and lipid peroxidation by ascorbic acid in the skin of mice Exposed to Fractionnated γ Radiation. Clin Chem Acta. 2003; 332:111-121.
- 53. Obara H, Harasawa R. L-Ascorbic acid enhances apoptosis in human gastric carcinoma n cell line Az-521 cells infected with mycoplasma hyorthinis .J Vet Med Sci. 2008; 70:11-15.
- 54. Oguteu A, Suludere Z, Kalender Y. Dichlorvos induced hepatotoxocity in rats and the protective effect of vitamin C and vitamin E. Env Toxicol Pharmacol. 2008; 26:355-361.
- 55. Nimse SB, Pal D. Free radicals, natural antioxidants, and their reaction mechanism. RSC Adv 5 2015; 27986-28006.
- 56. Hacioglu G, Senturk A, Ince I, Alver A. Assessment of oxidative stress parameters of brain derived neutrophil factor heterozygous Mice in acute stress model. Iran J Basic Med Sci. 2016; 19: 388-393.
- 57. Liu ZZ, Zhou T, Ziegler A, Dimitrion P, Zuo L. Neurodegenerative diseases: From molecular mechanism to clinical oxidation. Oxidative Med. Cellular. Logevity 2017; https://doi.org/10.115.
- 58. Oliveira GLS, Oliveira FFA, Freitas RM. Potential involvement of oxidative stress in induction of neurodegeneration disease: Actions, mechanisms and neutrotherapeutic potentials of natural antioxidants. Afr J Pharmacol. 2014; 8(25): 685-700.
- 59. Singh SK. Oxidative stress and neurodegeneration. J Cytol Histol 2017; 8:4 Dio: 10.4172/2157-7099.1000e119.
- 60. Klaunig JE, Kamendulis LM, Hocevar BA. Oxidative stress and oxidative damage in carcinogenesis. J Toxicol. Pathol. 2010; 38:96-109.
- 61. Reuter S, Gupta SC, Chaturvedi MM, Aggarwal BB. Oxidative stress, inflammation, and cancer: How are they Linked? Free Radic Bio Med. 2010; 49 (11): 1603-1616.
- 62. Toyokuni S. Molecular mechanisms of oxidative stress induced carcinogenesis: From epidemiology to oxygenomics. IUBMB Life. 2008; 60 (7): 441-447.
- 63. Niedowicz DM, Daleke DL. The role of oxidative in diabetic complication. Cell Biochem Biophysic. 2005; 43:289-330.
- 64. Ullah A, Khan A, Khan I. Diabetic mellitus and oxidative stress- A Concise review. Saudi Pharm J. 2016; 24:547-553.
- 65. Wright E, Scism-Bacon JL, Glass LC. Oxidative stress in Type 2 Diabates: The role of fasting and postprandial glycaemia. Int J Clin Pract. 2006; 60(3): 308-314.
- 66. Barrita SLJ, Sanchez SSM. Antioxidants role of ascorbic acid and the protective effects on chronic disease. In Tech Open Sci 2013; 450-484.
- 67. Ichim TE, Miner B, Bracia KT, Luna B, Hunning hake R, Mikirova NA et al. Intravenous ascorbic acid to prevent and treat cancer associated sepsis. J Translational Med. 2011; 9:25-38.
- 68. Yeon CH, Jung GC, Song KJ. Changes of terminal cancer patient health related quality of life after high dose vitamin C dministration. J Korean Med sci. 2007; 22: 7-11.
- 69. Coyle P, Kosnett MJ, Hinkins. Severe Lead Poisoning in the Plastics Industry; A Report of three Cases. Am J Indus Med. 2005; 47(2): 172-175.
- 70. Tsai M, Huang S, Cheng S. Lead Poisoning can be easily misdiagnosed as Acute Porphyria and non-specific abdominal pain. Case Report Emer Med. 2017; https://doi.org/10.1155/9050713.
- 71. Amadi CN, Igweze ZN, Oriskwe O.E. Heavy Metals in miscarriages and still birth in developing nations. J Middle East Fert Socie. 2017; 22(2):91-100.
- 72. Delgado CF, Ullery MA, Jordan M, Duclos C, Rajagopalan S, Scott K. Lead Exposure and Developmental Disabilities in Pre-school aged Children.www.JPHMP.COM 2017; 00:1-8.
- 73. Lipko M, Debski B. Mechanism of insulin like Effect of chromium (III) ions on glucose uptake in C2C2 Mouse Myotubes involves ROS formation. J Trace Elem Med Bio. 2018; 45:171-175.
- 74. Valko M, Moris H, Cronim MT. Metal toxicity and oxidative stress. J Curr Med Chem 2005; 12(10):1161-1208.
- 75. Khan AR, Awan FR. Metals in the pathogenesis of type 2 diabetes. J Diab Met Disord 2014; 13:16.
- 76. Price KD, Price CSC, Reynolds RD. Hyperglycemia induced ascorbic acid deficiency promotes endothelial dysfunction and development of atherosclerosis. J Arthero 2001; 158: 1-12.
- 77. Csanyi G, Miller FJ. Oxidative stress in cardiovascular disease. Int J Mol Sci 2014; 15:6002-6008.
- 78. Elahi MM, Kong YX, Matata BM. Oxidative stress as a mediator of cardiovascular disease. Oxidative Med Cellular Longev. 2009; 2 (5):259-269.
- 79. Lakshmi SVV, Padmaja G, Kuppusamy P, Kutala VK. Oxidative stress in cardiovascular disease. Indian J Biochem Biophys. 2009; 46:421-440.

Available online at https://saspublishers.com/journal/sjams/home

- 80. Mei Y, Thompson MD, Cohen RA, Tong XY. Autophagy and oxidative stress in cardiovascular disease. Biochemica et Biophysica Acta. 2015; 1852:243-251.
- 81. Otani H. Oxidative stress as pathogenesis of cardiovascular risk associated metabolic syndrome. J Antioxid Redox Signal. 2011; 15(11):1911-1926.
- 82. Li D, Mehta JL. Oxidized LDL, a critical factor in atherogenesis. Cardiovasc Res. 2005; 68:353-354.
- 83. Moser MA, Chun OK. Vitamin C and health: A review based on findings from epidemiologic studies. Int J Mol Sci. 2016; 17:1328.
- 84. Robitaille L, Mamer OA, Miller WH, Levine M, Assouline S, Melnychuck D et al. Oxalic acid excretion after intravenous ascorbic acid administration. Metabolism. 2009; 58(2); 263-269.
- 85. Baxmann AC, De OG, Mendonca C, Heilberg IP. Effect of vitamin C supplements on urinary oxalate and pH in calcium stone formation patients. Kidney Int. 2003; 63: 1066-71.
- 86. Chai W, Liebman M, Kynast-Gales S, Massey L. Oxalate absorption and endogenous oxalate synthesis from ascorbate in calcium ascorbate formers and non-stone formers. Am J Kidn Dis 2004; 44: 1060-9.
- 87. Chakraborthy A, Ramani P, Sherlin HJ, Premkumar P, Natesan A. Antioxidant and pro-oxidant activity of Vitamin C in oral environment. Indian J Dent Res. 2014; 25(4): 499-504.
- 88. Haovas Z, Sallem A, Zidi I, Hichri H, Mzali I, Mehdi M. Hepatotoxic effect of lead acetate in rats: Hepatopathological and cytotoxic Studies. J Cytol Histol. 2014; 5:256.
- 89. Hermane DS, Geraldine M, Venkatesh T. Influence of minerals on lead induced alteration in liver function in rats exposed to long term lead exposure. J Hazard. 2009; 166(2):1410-1414.
- Omotosho BR, Abiodun AA, Ijomone MN, Adewoye SO. Lead induced damaged on hepatocytes and hepatic reticular fibre in rats: Protective role of aqeous extract of *Moringa oleifera* leaves (Lam). J Biolog Sci Med. 2015; 3:27-35.
- 91. Ozsoy SY, Ozsoy B, Ozyildiz Z, Aytekan. Protective effect of L-Carnitine on experimental lead toxicity in rats: A Clinical, histopathological and immunohistochemical study. J Bioteh Histochem 2011; 86(6):436-443.
- 92. Shalan MG, Mostafa MS, Hassouna MM, Hassab SE, El-Refaie A. Amelioration of lead toxicity on lead toxicity on rat liver with Vitamin C and Silymarin supplementation. Toxicology 2005; 206 (1):1-5.
- 93. Aziz FM, Maulood IM, Chawsheen MAH. Effect of melatonin, Vitamin C and E alone or in combination on lead induced injury in liver and kidney organs of albino Rats. J Pharm 2013; 2(5):13-18.
- 94. El-Tohamy MM, El-Natt W.S. Effect of antioxidant on lead induced oxidative damage and reproductive dysfunction in male rabbits. J Am Sci. 2010; 6(11):613-622.
- 95. Rajamanichan V, Muthuswamy N. Effect of heavy metals induced toxicity on metabolic biomarkers in common Carp (*Cyprinus carpiol L*). Maejo Internat J Sci Techno. 2008; 2(1):192-200.
- 96. Dorostghoal M, Dezfoolian A, Sorooshnia F. Effect of maternal lead acetate exposure during lactation and postnatal development of testis in offspring Wister rats. Iranian J Basic Med Sci. 2010; 14(2):122-131.
- 97. Vigeh M, Smith DR, Hsu P. How does lead induce male infertility? Iranian J Repro Med 2011; 9 (1):1-8.
- 98. Lewis MW, Pitts DK. Inorganic lead exposure activates Striatal cFOS expression at lower blood levels and inhibits amphetamine induced Cfos expression at higher blood levels. J Pharmacol. 2004; 310(2):815-820.
- 99. Morga S, Sharma R, Qureshi N. Effect of maternal lead acetate exposure on prenatal development of Swiss albino mice. Asian J Env Sci. 2009; 4(2):216-220.
- 100. Hertz PI. The evidence that lead increases the risk for spontaneous abortion. Am J Indust Med 2000; 38:300-309.
- 101. Biswas NM, Gosh P. Effect of lead on male gonadal in albino rats. J Kathmandu Uni Medic 2004; 2(1):43-46.
- 102. Ahmad I, Sabir M, Yasin KF. Study of the effects of lead poisoning on the testes in albino rats. J Med Res. 2003; 42:1-9.
- 103. Fahin MA, Tariq S, Adeghate E. Vitamin E modifies the ultrastructure of testis and epididymis in mice exposed to lead intoxication. Ann Anat. 2013; 195:272-277.
- 104. Archarya UR, Rathore RM, Mishra M. Role of vitamin C on lead acetate induced spermatogenesis in Swiss mice. J Env Toxicol Pharmacol. 2003; 13:9-14.
- 105. Apaydin FG, Kalender S, Bas H, Demir F, Kalender Y. Lead nitrate induced testicular toxicity in diabetic and non-diabetic rats. Protective role of sodium selenite. Brazilian archives. Biol Technol. 2015; 58(1):68-74.
- 106. Turner TT, Lysiak JJ. Oxidative stress: A Common factor in testicular dysfunction. J Androl. 2008; 29 (3):488-498.
- 107. Hsu PC, Liu MY, Hsu CC, Chen LY, Guo LY. Effects of vitamin E and/or C on reactive oxygen species (ROS) related lead toxicity in the rats' sperm. J Toxicol. 1998; 128:169-179.
- 108. Shan G, Tang T, Zhang X. The protective effect of ascorbic acid and thiamine supplementation against damage caused by lead in the testes of mice. J Tech Med Sci. 2009; 29(1):68-72.
- Shabanian S, Farahbod F, Rafiean M, Ganj F, Adib A. The effects of vitamin C on sperm quality parameters in laboratory rats following long term exposure to cyclophosphamide J Adv Pharmaceu Tech Res. 2017; 8 (2): 73-79.
- 110. Agawal A, Sekhon LH. The role of antioxidant therapy in the treatment of male infertility. Hum Fert. 2010; 13:217-225.

- 111. Mamoun MA, Rizvi SMTA, Qazi SM. Reduction of testicular volume of albino rats in lead induced toxicity and reversal of it with high dosage of vitamin C. Med Forum. 2015; 26 (2): 15-19.
- 112. Sharma DN. Ascorbic acid protects testicular oxidative stress and spermatozoa deformation in Male Swiss mice exposed to lead acetate. Universal J Env Res Tech. 2013; 3(1):86-92.
- 113. Shafai AE, Zohdy N, Mulla KE, Hassan M, Morad N. Electron microscopic study of the toxic effects of prolonged lead exposure on the seminiferous tubles of albino rats and the possible protective effect of ascorbic acid. Fd Chem Toxicol. 2010; 49: 734-743.
- 114. Sonmez M, Turk G, Yuce A. The Effect of ascorbic acid supplementation on sperm quality, lipid peroxidation, and testosterone levels of male Wister Rats. Theriogenology. 2005; 63:2063-2072.
- 115. Azari O, Gholipour H, Kheirandish R, Babaei H, Emadil L. Study of the protective effects of vitamin C on testicular tissue following experimental unilateral cryptochidsm in rats. Angrologia. 2014; 46 (5):495-503.
- 116. Fraga CG, Motchinic PA, Shigenaga MK, Helbock HJ, Jacob RA, Ames BN. Ascorbic acid protects against endogenous oxidative DNA damage in hum sperm. Proct Natl Acad Sci. 1991; 188:11003-11006.
- 117. Wang C, Zhang Y, Liang J, Shan G, Wang Y, Shi Q. Impacts of ascorbic acid and thiamine supplementation at different concentrations on lead toxicity in testes. Clin Chem. 2006; 370:82-88.
- 118. Allouche L, Hamadouche M, Touabti A, Khennouf S. Effect of long term exposure to low or moderate lead concentrations on growth, lipid profile and liver function in Albino Rats. Adv Bio Res. 2011; 5(6):339-347.
- 119. Ademuyiwa O, Ugbaja RN, Idumebor F, Adebawa O. Plasma lipid profiles and risk of cardiovascular disease in Occupational lead exposure in Abeokuta, Nigeria. Lipids in health and diseases. 2005; 4:19.
- 120. Heo Y, Lee BK, Ahn KD, Lawrence DA. Serum IgE elevation correlate with Blood lead levels in battery manufacturing workers. Human Expo Toxicol. 2004; 23:209-213.
- 121. Newairy AA, Abdou HM. Protective Role of flax lignans against lead acetate induced oxidative damage and hyperlipidaemia in Rats. Fd Chem Toxicol 2009); 47(4):813-818.
- 122. McRae MP. Vitamin C supplementation lowers serum low density lipoprotein cholesterol, triglyceride: a Metaanalysis of 130 Randomised Control trials. J Chiropractic Med. 2008; 7(2):48-58.
- 123. Dominiczak MH, McNamara JR. The System of cardiovascular prevention. In Handbook of Lipoprotein Testing. Eds. Rifai N, Narmnick and GR and Dominiczak MH. American Association for Clinical Chemistry Inc. Washington 2000; Pp 103.
- Murphy JG, Lioyd MA. Mayo clinic cardiology: Concise Textbook. Moyo Clinic Science Press, Rochester. 2007; Pp 715.
- 125. Ugbaja RN, Okonkwo BO, Omoniyi DA. Lead induced dyslipidemia. The comparative effects of ascorbate and chelation Therapy. Afri J Biotchn. 2013; 12 (15):1845-1852.
- 126. Etteng MU, Ibekwe HA, Amatey TE, Bassey BJ, Oboh FU, Owu DU. Effect of vitamin C on serum lipids and electrolyte profile of albino Wistar rats. Nig J Phys Sci. 2006; 21 (2): 15-19.
- 127. Deicher R, Zia F, Bieglmer C, Schillinger M, Horl WH. Low total vitamin C plasma level is a risk factor for cardiovascular morbidity and mortality in hemodialysis patients. J Amer Society Nephro. 2005; 16:1811-1818.
- Alexa ID, Mihalache I, Panaghiu L, Palade F. Chronic lead poisoning- A forgotten cause of anemia. Rev Med Chir Soc Med Nat lasi. 2002; 106(4):825-8.
- 129. Noori MM, Heidari Z, Mahmoudzadeh SH, Barbarestani M. Effect of chronic lead acetate intoxication on blood indices of male adult rat. J Daru Pharmacol. 2003; 11(4):145-51.
- Sivaprasad R, Nagaraj M, Varalakshmi P. Combined efficacies of lipoic acid and meso-2,3-dimecarptosuccinic acid on lead induced erythrocytes membrane lipid peroxidation and antioxidant status in rats. Hum Expos Toxicol. 2003; 22:83-192.
- 131. Gonzalez MJ, Mirande-Massari JR. New insights on vitamin C and cancer. Springer Briefs in Cancer Research 2014; Dio10.1007/978-1-4939-1890-4.
- 132. Ottoboni F, Ottoboni A. Ascorbic acid and the immune system. J Orthomolec Med. 2005; 20 (3): 179-183.
- Othman AI, Alsharawy S, Elmissiry. Role of melatonin in ameliorating lead induced haematotoxicity. J Pharmacol Res. 2004; 50(3):301-7.
- 134. Pratinidhi SA, Patil JA, Bahera M, Patil, Gbadage DP, Pratinidhi A. Effects of blood lead level on biochemical and haematological parameters in children with neurological diseases in Western Maharashtra, India. J Bas Clin Physiol Pharmacol. 2014; 25(2): 229-233.
- 135. Dey PM, Burger J, Gochfeld M, Reuhl KR. Developmental lead exposure disturbs expression of synaptic neural cell adhesion molecule in herring gull brains. Toxicology. 2005; 146(2-3), 137-147.
- 136. Gurer-Orhan H, Sabir HU, Ozgunes H. Correlation between clinical indicators of lead poisoning and oxidative stress parameters in control and lead exposed workers. J Toxicol 2004; 195(2): 147-54.
- 137. Marchetti C. Molecular targets of Lead in brain neurotoxicity. Neuro Toxic Res. 2003; 5(3): 221-35.
- 138. Nour-Eddine D, Miloud S, Abdelkader A. Effect of lead exposure on doperminergic transmission in the rat's brain. Toxicol 2005; 207(3): 363-8.
- 139. Shawky SM, Ramadan SGA, Orabi SH. Hemato-biochemical, behavioral and neurological effects of vitamin C administration against lead exposure in Mice. Int J Adv Res. 2014; 2(12): 418-429.

Available online at https://saspublishers.com/journal/sjams/home

- 140. Sadeghi A, Bideskan AE, Allipour F, Fazel A, Haghir H. The effect of ascorbic acid and Garlic acid administration on lead induced neural damage in rat offspring hippocampus. Iranian J Bas Med Sci 2013; 16(2):157-164.
- 141. Harrison FE, May JM. Vitamin C Function in the Brain: Vital bole of the Ascorbate transporters SVCT<sub>2</sub>. Free Radic Bio Med. 2009; 46 (6): 719-30.
- 142. Jalil EC, Akundi RS, Bhatia HS, Lieb K, Appel K, Munoz E, Huh M. Ascorbic acid enhances the inhibitory effect of aspirin on neuronal cyclooxygenase-2- mediated prostaglandin E<sub>2</sub> production. J Neuroimmunol. 2006; 174 (2): 39-5.
- 143. Salehi I, Soleiman MS, Pourjafar M, Moravej FG, Asi SS. Vitamin C can alter lead induced passive avoidance learning impairment in rats. Anatomical Sci. 2015; 12(3): 137-40.
- 144. Musa SA, Omoniye IM, Hamman WO, Ibegbu AO, Umana UE. Preventive activity of ascorbic acid on lead acetate induced cerebellar damage in adult Wister rats. Med Health Sci. J 2014; 13: 99-104.