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Physicochemical and Phytochemical Standardization of Siddha Herbal Drug Formulation- Parangichakkai Chooranam

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Abstract

Original Research Article

The Siddha system of medicine is capable of treating all types of skin diseases. This trial medicine Parangichakkai chooranam is the sasthric preparation. The aim of the present study is investigating the phytochemical and physiochemical analysis of the trial drug Parangichakkai chooranam. Phytochemical analysis, heavy metal analysis and physiochemical parameters such as ash values, extract values, loss of drying were determined as protocol. The physiochemical parameters such as the total as value was found to be 14.3 ± 1.054 , insoluble ash value 0.066 ± 0.041 , loss of drying 11.5 ± 0.9 and water-soluble extract 5.213 ± 0.9 , alcohol soluble extract found to be 13.8 ± 2.553 . the phytochemical analysis of different extracts gave positive test for Alkaloids, flavonoids, steroids, coumarin, phenol, tannin, protein, sugar. The heavy metal analysis showed the presence of heavy metal mercury at 0.027ppm, which may be less than the recommended limit. The results obtained indicate that the drug is of standard quality and can be used as reference standard.

Keywords: Parangichakkai chooranam, Siddha medicine, Physicochemical analysis, Herbal drug.

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INTRODUCTION

A siddha system of medicine is the oldest holistic management system with meticulously documented medicines and being practiced by large population in south India. Herbal traditional medicines have gained considerable momentum worldwide during the past decade and play paramount role in health care programme especially in developing countries. According to W.H.O nearly 80% of population of developing countries rely on traditional medicines for most of their ailments. Ancient siddha literatures are well provided with reference on the use of herbs with medicinal properties. Balakarappan has been described in siddha literature as the diseases occurring in children. According to the siddha textbook 'Kuzhandhai Maruthuvam (Balavagadam) [1]. Atopic dermatitis in infants and children the rash usually occurs on the scalp, back of the knees and cheeks, elbows. It is character by dry, itchy, red and cracked skin, which can sometimes ooze fluid and bleed. Parangichakkai chooranam comprises of 21 herbal ingredients like Smilax china, Zingiber officinale, Piper longum, Elettaria cardamomum, Embelia ribes, Cinnamomum verum, Carum copiticum, Hyoscyamus niger, Plumbago zeylanica, Alpinia officinarum, Piper longum, Alpinia galangal, Clerodendrum serratum, Coriandrum sativum, Cuminum cyminum, Nigella sativa, Glycyrrhiza glabra, Vettiveria zizanioides, Plectranthus vettiveroides, Cyperus rotundus, Kaempferia galangal. The herbal ingredients of parangichakkai chooranam possess antiinflammatory, anti-microbial, anti-bacterial, antiimmune and allergic activity. which have the tendency to cure skin diseases. The present study is to investigate the physiochemical and phytochemical and heavy metal analysis of the trial drug parangichakkai chooranam sasthric preparation which is mentioned in siddha text book of chikicha rathna deepam.

Ingridients of parangichakkai chooranam [2, 3]

- 1. Parangichakkai (Smilax china) (105 gm) [4]
- 2. Sukku (Zingiber officinale) (12.6gm)
- 3. Thippli (Piper longum) (12.6gm)
- 4. Elam (Elettaria cardamomum) (12.6gm)
- 5. Vaaivilangam (Embelia ribes) (12.6gm)
- 6. Sannalavangapattai (Cinnamomum verum) (12.6gm)

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- 7. Omam (Carum copticum) (12.6gm)
- 8. Kurosani omam (Hyoscyamus niger) (12.6gm)
- 9. Chitramoola verpattai (Plumbago zeylanica) (12.6gm)
- 10. Chitrarathai (Alpinia officinarum) (12.6gm)
- 11. Thippliver (Piper longum) (12.6gm)
- 12. Perarathai (Alpinia galanga) (12.6gm)
- 13. Sirutheku (Clerodendrum serratum) (12.6gm)
- 14. Dhaniya (Coriandrum sativum) (12.6gm)
- 15. Seeragam (Cuminum cyminum) (12.6gm)
- 16. Karunjeeragam (Nigella sativa) (12.6gm)
- 17. Adhimaduram (Glycyrrhiza glabra) (12.6gm)
- 18. Vettiver (Vetiveria zizanioides) (12.6gm)
- 19. Vilamichuver (Plectranthus vettiveroides) (12.6gm)
- 20. Muthakasu (Cyperus rotundus) (12.6gm)
- 21. Kichilikizangu (Kaempferia galanga) (12.6gm)

METHOD OF PREPARATION

All the above ingredients are purified as per book Sarakku suthi muraigal and are grinded separately as fine powder. Then the powder is mixed together with equal amount of white sugar. Prepared medicine is stored in clean and dry glass container.

Dosage

5-6 years - 1.3 gm 7- 12 years - 2 gm, twice a day, after food Vehicle: Ghee

Duration of Treatment: 45 days

Physiocochemical Analysis [5, 6]:

Table-1: Sample Description

State	Solid
Nature	Coarse powder
Odor	Aromatic
Touch	Slightly moistened
Flow Property	Non free flowing
Appearance	Pale Greenish brown

Percentage Loss on Drying

Test drug was accurately weighed in evaporating dish. The sample was dried at 105° C for 5 hours and then weighed.

Determination of Total Ash

Test drug was accurately weighed in silica dish and incinerated at the furnace with the temperature of 400 °C until it turns white in color which indicates absence of carbon. Percentage of total ash will be calculated with reference to the weight of air-dried drug.

Determination of Acid Insoluble Ash

The ash obtained by total ash test was boiled with 25 ml of dilute hydrochloric acid for 6 mins. Then the insoluble matter was collected in crucible and washed with hot water and ignited to constant weight. Percentage of acid insoluble ash was calculated with reference to the weight of air-dried ash.

Determination of Alcohol Soluble Extractive

Test sample was macerated with 100 ml of alcohol in a closed flask for twenty-four hours, shaking frequently during six hours and allowing it to stand for eighteen hours. Filter rapidly, taking precautions against loss of solvent, evaporate 25 ml of the filtrate in a tared flat-bottomed shallow dish, and dry at 105°C, to constant weight and weigh. Calculate the percentage of alcohol-soluble extractive with reference to the air-dried drug.

Determination of Water-Soluble Extractive

Test sample was macerated with 100 ml of chloroform water in a closed flask for twenty-four hours, shaking frequently during six hours and allowing it to stand for eighteen hours. Filter rapidly, taking precautions against loss of solvent, evaporate 25 ml of the filtrate to dryness in a tared flat-bottomed shallow dish, and dry at 105°C, to constant weight and weigh. Calculate the percentage of water-soluble extractive with reference to the air-dried drug.

Tuble 2. That Test Teport						
S. No	Parameter	Mean (n=3) SD				
1.	Loss on Drying at 105 °C (%)	11.5 ± 0.9				
2.	Total Ash (%)	14.3 ± 1.054				
3.	Acid insoluble Ash (%)	0.066 ± 0.041				
4.	Water soluble Extractive (%)	5.213 ± 0.91				
5.	Alcohol Soluble Extractive (%)	13.8 ± 2.553				

Table-2: Final Test report

Sterility Test by Pour Plate Method Objective

The pour plate techniques were adopted to determine the sterility of the product. Contaminated / un sterile sample (formulation) when come in contact with the nutrition rich medium it promotes the growth of the organism and after stipulated period of incubation the growth of the organism was identified by characteristic pattern of colonies. The colonies are referred to as Colony Forming Units (CFUs).

METHODOLOGY

Test sample was inoculated in sterile petri dish to which about 15 mL of molten agar 45°C were added. Agar and sample were mixed thoroughly by tilting and swirling the dish. Agar was kept undisturbed for 10 minutes. Plates were then inverted and incubated at 37° C for 24-48 hours and further extended for 72 hours for fungal growth observation. Grown colonies of organism was then counted and calculated for CFU.

RESULT

No growth / colonies were observed in any of the plates inoculates with the test sample. **Test for Specific Pathogen**

Methodology

Test sample was directly inoculated in to the specific pathogen medium (EMB, DCC, Mannitol, Cetrimide) by pour plate method. The plates were incubated at 37°C for 24 - 72h for observation. Presence of specific pathogen identified by their characteristic color with respect to pattern of colony formation in each differential media.

Table-5. Detail of Specific Median and then aboreviation				
Organism	Abbreviation	Medium		
E-coli	EC	EMB Agar		
Salmonella	SA	Deoxycholate agar		
Staphylococcus Aureus	ST	Mannitol salt agar		
Pseudomonas Aeruginosa	PS	Cetrimide Agar		

Table-3: Detail of Specific Medium and their abbreviation

Staphylococcus Auleus	51	Mainin	of salt agai	
Pseudomonas Aeruginosa	PS	Cetrim	ide Agar	
Table-4: Result of Specific pathogen				
Organism	Spec	ification	Result	
E-coli	Abse	ent	Absent	
Salmonella	Abse	ent	Absent	

Absent

Absent

No growth / colonies were observed in any of the plates inoculated with the test sample.

Staphylococcus Aureus

Pseudomonas Aeruginosa

Heavy Metal Analysis by AAS

Standard: Hg, As, Pb and Cd - Sigma

Methodology

Atomic Absorption Spectrometry (AAS) is a very common and reliable technique for detecting metals and metalloids in environmental samples. The total heavy metal content of the sample was performed by Atomic Absorption Spectrometry (AAS) Model AA 240 Series. In order to determination the heavy metals such as mercury, arsenic, lead and cadmium concentrations in the test item.

Absent

Absent

Sample Digestion

Test sample was digested with 1mol/L HCl for determination of arsenic and mercury. Similarly, for the determination of lead and cadmium the sample were digested with 1mol/L of HNO3.

Standard Reparation

As & Hg- 100 ppm sample in 1mol/L HCl Cd & Pb- 100 ppm sample in 1mol/L HNO3

Table-5: Heavy metal analysis					
Name of the Heavy Metal	Absorption Max Λ max	Result Analysis	Maximum Limit		
Mercury	253.7 nm	0.027	1 ppm		
Lead	217.0 nm	BDL	10 ppm		
Arsenic	193.7 nm	BDL	3 ppm		
Cadmium	228.8 nm	BDL	0.3 ppm		

Particle Size Determination by Microscopic Method [8]

Methodology

Particle size determination was carried out by optical microscopic method. In which the sample were dissolved in the sterile distilled water (app 1/100th dilution). Diluted sample were mounted on the slide and fixed with stage of appropriate location. Light microscopic image was drawn with scale micrometer to arrive at the average particle size. Minimum 30 observations were made to ascertain the mean average particle size of the sample.



Fig-1: Microscopic Observation of Particle Size for the sample PC

REPORT

Microscopic observation of the particle size analysis reveals that the average particle size of the sample PC was found to be 110.6 ± 41.54 µm further the sample PC has particle with the size range of lowest 46 µm to highest 187 µm

Phytochemical analysis of Parangichakkai Chooranam [7]

Test for alkaloids

Mayer's Test: To the test sample, 2ml of mayer's reagent was added, a dull white precipitate reveals the presence of alkaloids.

Test for coumarins

To the test sample, 1 ml of 10% sodium hydroxide was added. The presence of coumarins was indicated by the formation of yellow color.

Test for saponins

To the test sample, 5 ml of water was added and the tube was shaken vigorously. Copious lather formation indicates the presence of Saponins

Test for tannins

To the test sample, ferric chloride was added, formation of a dark blue or greenish black color showed the presence of tannins.

Test for glycosides- Borntrager's Test

Test drug was hydrolyzed with concentrated hydrochloric acid for 2 hours on a water bath, filtered and the hydrolysate was subjected to the following tests. To 2 ml of filtered hydrolysate, 3 ml of chloroform was added and shaken, chloroform layer was separated and 10% ammonia solution was added to it. Pink color indicates the presence of glycosides.

Test for flavonoids:

To the test sample about 5 ml of dilute ammonia solution was added followed by addition of few drops of conc. Sulfuric acid. Appearance of yellow color indicates the presence of Flavonoids. Test for phenols:

Lead acetate test: To the test sample; 3 ml of 10% lead acetate solution was added. A bulky white precipitate indicates the presence of phenolic compounds.

Test for steroids:

To the test sample, 2ml of chloroform was added with few drops of conc. Sulphuric acid (3ml), and shaked well. The upper layer in the test tube was turned to red and sulphuric acid layer showed yellow with green fluorescence. It showed the presence of steroids.

Triterpenoids

Liebermann–Burchard test: To the chloroform solution, few drops of acetic anhydride was added and mixed well. 1 ml concentrated sulphuric acid was added from the sides of the test tube, appearance of red ring indicates the presence of triterpenoids.

Test for Cyanins

Anthocyanin:

To the test sample, 1 ml of 2N sodium hydroxide was added and heated for 5 min at 100°C. Formation of bluish green color indicates the presence of anthocyanin.

Test for Carbohydrates - Benedict's test

To the test sample about 0.5 ml of Benedict's reagent was added. The mixture was heated on a boiling water bath for 2 minutes. A characteristic colored precipitate indicates the presence of sugar.

Proteins (Biuret Test)

To extracts 1% solution of copper sulphate was added followed by 5% solution of sodium hydroxide, formation of violet purple color indicates the presence of proteins.

Results of phytochemical analysis:

The phytochemical analysis showed the presence of alkaloids, flavonoids, steroids, coumarin, phenol, tannin, protein, sugar.



Fig-2: Qualitative Phytochemical Investigation

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Fig-3: Test for Alkaloids, Flavonoids, Glycosides, Steroids and Triterpenoids



Fig-4: Test for Coumarin, Phenol, Tanins, Saponin, Proteins

Biochemical analysis Parangichakkai Chooranam Preparation of Extract:

5gm of sample was taken in a 250ml clean beaker and added with 50ml of distilled water. Then it was boiled well for about 10 minutes. Then it was cooled and filtered in a 100ml volumetric flask and made up to 100ml with distilled water. This preparation was used for the qualitative analysis of acidic/basic radicals and biochemical constituents in it.

Procedure:

Test for Silicate

A 2ml of the sample was shaken well with distilled water.

Action of Heat:

A 2ml of the sample was taken in a dry test tube and heated gently at first and then strong.

Ash Test:

A filter paper was soaked into a mixture of extract and dil. cobalt nitrate solution and introduced into the Bunsen flame and ignited.

Test for Acid Radicals Test for Sulphate:

2ml of the above prepared extract was taken in a test tube to this added 2ml of 4% dil ammonium oxalate solution.

Test for chloride:

2ml of the above prepared extracts was added with 2ml of dil. HCl until the effervescence ceases off.

Test for Phosphate:

2ml of the extract were treated with 2ml of dil. ammonium molybdate solution and 2ml of con. HNo₃.

Test for carbonate:

2ml of the extract was treated with 2ml of dil. magnesium sulphate solution.

Test for Nitrate:

lgm of the extract was heated with copper turning and concentrated H_2So_4 and viewed the test tube vertically down.

Test for Basic radicals

Test for lead:

2ml of the extract was added with 2ml of dil. potassium iodine solution.

Test for copper:

One pinch (25mg) of extract was made into paste with con. HCl in a watch glass and introduced into the non-luminous part of the flame.

Test for Aluminum:

To the 2ml of extract dil. sodium hydroxide was added in 5 drops to excess.

Test for Iron:

a. To the 2ml of extract was added 2ml of dil. ammonium solution

b. To the 2ml of extract 2ml of thiocyanate solution and 2ml of con HNo_3 is added.

Test for Zinc:

To 2ml of the extract dil. sodium hydroxide solution was added in 5 drops to excess and dil. ammonium chloride was added.

Test for Calcium:

To 2ml of the extract was added 2ml of 4% dil. ammonium oxalate solution.

Test for Magnesium:

To 2ml of extract dil. sodium hydroxide solution was added in drops to excess.

Test for Ammonium:

To 2ml of extract 1 ml of Nessler's reagent and excess of dil. sodium hydroxide solution were added.

Test for Potassium:

A pinch (25mg) of extract was treated off with 2ml of dil. sodium nitrite solution and then treated with 2ml of dil. cobalt nitrate in 30% dil. glacial acetic acid.

Test for Sodium:

2 pinches (50mg) of the extract was made into paste by using HCl and introduced into the blue flame of Bunsen burner.

Test for Mercury:

2ml of the extract was treated with 2ml of dil. sodium hydroxide solution.

Test for Arsenic:

2ml of the extract was treated with 2ml of dil. sodium hydroxide solution

Miscellaneous

Test for Starch:

2ml of extract was treated with weak dil. Iodine solution.

Test for Reducing Sugar:

5ml of Benedict's qualitative solution was taken in a test tube and allowed to boil for 2 minutes and added 8 to 10 drops of the extract was added and again boiled it for 2 minutes. The color changes were noted.

Test for the Alkaloids:

a) 2ml of the extract was treated with 2ml of dil. potassium Iodide solution.

b) 2ml of the extract was treated with 2ml of dil. picric acid.

c) 2ml of the extract was treated with 2ml of dil. phosphotungstic acid.

Test for Tannic Acid:

2ml of extract was treated with 2ml of dil. ferric chloride solution.

Test for Unsaturated Compound:

To the 2ml of extract, 2ml of dil. Potassium permanganate solution was added.

Test for Amino Acid:

2 drops of the extract were placed on a filter paper and dried well. 20ml of Burette reagent is added.

Test for Type of Compound:

2ml of the extract was treated with 2 ml of dil. ferric chloride solution.

DISCUSSION AND CONCLUSION

From the current study of Preclinical standardization of Parangichakkai chooranam which is mentioned in siddha texts shows that the drug parangichakkai chooranam was fine powder pale greenish brown in color with aromatic, slightly bitter and sweet taste. The drug size has particle size with the range of lowest 46 µm to highest 187µm. the loss on drying indicates the moister content of the drug was determined as 11.5±0.9%. The total ash was found to be 14.3±1.054% which indicates inorganic content of the drug. The water-soluble extract was calculated as 5.213±0.91% and the value of acid insoluble as was found to be 0.066±0.041%. the minimal level acid insoluble ash shows the less inorganic residue and alcohol soluble extract was found to be $13.8\pm2.553\%$. in this trial drug water soluble extract was found to be more than the alcohol soluble extract. This increased water soluble extract may produce immediate drug action. The phytochemical analysis shows the drug has high polar secondary metabolites like alkaloids, flavonoids, steroids, coumarin, phenol, tannin, protein and sugar. In heavy metal analysis of mercury was 0.027 and lead, arsenic, cadmium was present within the below detectable limit there by ensures its safe usage. This study also relevels that the chooranam is sterile and free of bacteria, fungi and specific pathogen like E-coli, salmonella, staphylococcus aureus, pseudomonas aeruginosa. The Biochemical analysis of parangichakkai chooranam revealed the presence of sulphate, chloride, phosphate, carbonate, iron, arsenic and alkaloids. The above the analysis showed the sample description, physicochemical, phytochemicals and heavy metals. As a result, parangipattai chooranam was proved its safety over the defined standardization

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method. The results obtained could be utilized as reference for developing standard formulation.

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