Scholars Journal of Applied Medical Sciences

Abbreviated Key Title: Sch J App Med Sci ISSN 2347-954X (Print) | ISSN 2320-6691 (Online) Journal homepage: https://saspublishers.com **3** OPEN ACCESS

Medical Sciences

Presence of Helicobacter Pylori in Lamina Propria and its Muscular Invasion in Oral Squamous Cell Carcinoma

Dr. Kavita Gupta^{1*}, Dr. Leeky Mohanty¹, Dr. Chaitanya. N. Babu¹

¹RGUGHS, The Oxford Dental College, Bangalore

DOI: https://doi.org/10.36347/sjams.2025.v13i12.010 | **Received:** 18.10.2025 | **Accepted:** 13.12.2025 | **Published:** 17.12.2025

*Corresponding author: Dr. Kavita Gupta RGUGHS, The Oxford Dental College, Bangalore

Abstract Original Research Article

Background and Objectives: Head and neck carcinomas are the biological heterogeneous group of cancers, of which oral carcinoma is the most common ninty one percent of oral cancers are squamous cell carcinomas originating from the mucosal epithelium. Oral carcinogenesis is a multifactorial process where multiple risk factors are involved. A connection between bacterial infection and carcinogenesis is convincing and is creating lots of interest now a days. Helicobacter pylori (H.pylori) are flagellated, gram negative, spiral, microaerophilic bacteria and deemed by the World Health Organization and the International Agency for Research on Cancer as a class I human carcinogen. In addition to its role in causing gastritis, gastric ulcer, duodenal ulcer and adenocarcinoma of stomach, H.pylori has been found in the oral cavity in patients with halitosis, apthous stomatitis and periodontal diseases. However, there are limited studies available in literature of H.pylori in oral squamous cell carcinoma (OSCC) and its possible role in oral carcinogenesis. Therefore, the aim of this study is to detect and establish a relationship between H.pylori and OSCC. Materials and Methods: Thirty paraffin embedded tissue blocks of clinically diagnosed and histopathologically confirmed cases of OSCC and ten of normal buccal mucosa were sectioned and stained with Hematoxylin and Eosin. The one subsequent serial section is immunohistochemically stained with anti-human helicobacter pylori antibody. The presence of H.pylori is assessed and any association between presence of H.pylori and OSCC was analyzed. The study data was analysed using SPSS (Statistical Package for Social Sciences) software V.22, IBM, Corp. Statistical Analysis was done using Chi Square test and Mann Whitney U test. The level of significance [P-Value] was set at P<0.01. **Results**: Chi Square test was used to find the association between the prevalence of H.Pylori between OSCC & Control group and it was found to be stastically significant (at P<0.001). Chi Square test showed significant differences between H. pylori positivity and different tissue types i.e in lamina propria and in the muscle layer of OSCC. Conclusion: There is a positive relation between the presence of H. pylori and OSCC. Therefore, early detection and eradication of H. pylori in high-risk patients are suggested.

Keywords: Oral squamous cell carcinoma, Helicobacter pylori, Immunohistochemistry.

Copyright © 2025 The Author(s): This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International License (CC BY-NC 4.0) which permits unrestricted use, distribution, and reproduction in any medium for non-commercial use provided the original author and source are credited.

Introduction

H.pylori is a gram negative, motile, microaerophillic, spiral shaped bacteria colonizing the human gastric mucosa, it affects almost half of world's population. It is designated as a type I carcinogen by World Health Organization with approximately affecting 80% of Indian adults [1]. H.pylori is found to be associated with the development of gastric adenocarcinoma, gastritis, gastric ulcer, duodenal ulcer but its presence is also detected in patients with halitosis, apthous stomatitis and PD2 Oral squamous cell carcinoma is the most common malignancy of the oral cavity and is particularly common in developing world like India. The etiology of OSCC is found to be multifactorial with various factors contributing to its pathogenesis. Various studies have

revealed an emerging role of bacteria in development of OSCC however they generally failed to conclusively establish any association. In this study we tried to find out the presence of H. pylori in OSCC and to establish if any association exist between them mmunohistochemically.

MATERIALS AND METHOD

Thirty formalin fixed, paraffin embedded biopsy specimens with clinically and pathologically confirmed OSCC cases and 10 cases of normal oral mucosa (NOM) samples were obtained from the Department of Oral and maxillofacial Pathology. One endoscopic biopsie of patient positive for H.pylori is taken as positive control. The study has been approved

Citation: Kavita Gupta, Leeky Mohanty, Chaitanya. N. Babu. Presence of Helicobacter Pylori in Lamina Propria and its Muscular Invasion in Oral Squamous Cell Carcinoma. Sch J App Med Sci, 2025 Dec 13(12): 2008-2015.

by our institutional ethical committee review board (Ref No.218 /2014-2015).

Hematoxylin and eosin staining

Formalin fixed, paraffin embedded specimens were cut into 4um sections and were stained with hematoxylin and eosin for histologic confirmation of clinical diagnosis and to detect H.pylori presence in OSCC. Additional sequential sections were prepared for immunohistochemical studies.

Immunohistochemistry protocol

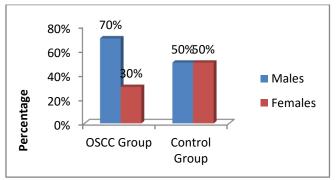
All the 30 cases of OSCC and 10 cases of normal buccal mucosa were available for high quality immunohistochemical staining. Immunohistochemical staining was performed on 4µm thick sections. All the procedures were performed at room temperature. The sections were deparaffinised through a series of xylene baths and rehydrated in graded concentrations of alcohol. Tissue sections were treated with 3% hydrogen peroxide to block endogenous peroxidase activity. Antigen retrieval was carried out by microwave with 0.01 M sodium citrate buffer solution for tree cycles of 800 W for 5 min twice and 200W for 14 min and later was subjected to two washes of tris buffer solution for 5 min each. Sections were then incubated with ready to use primary antibody (rabbit polyclonal to H.pylori, DAKO, Bengaluru, India). After washing with tris buffer solution, the sections were then incubated for 30 min with anti-mouse secondary antibody and visualized using 3, 3'-diaminobenzidine (DAB) chromogen. Section then visualized under microscope (4X, 10X and 40X) magnification and were assessed for the cytotoxic associated gene A expression on the outer membrane of H.pylori.

Statistical analysis

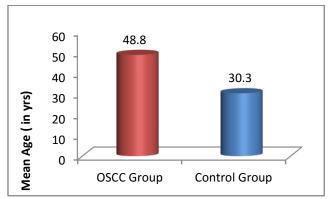
Helicobacter pylori presence was looked in the connective tissue and any correlation was assessed and they were analysed as present or absent. The data obtained were tabulated. Statistical values were analyzed using Chi-square test and Mann Whitney U test. A p-value of 0.05 or less was considered statistically significant.

RESULTS

In our study, the age of the OSCC patients ranged from 35 to 68 years and patients of normal buccal mucosa ranged from 23 to 42 years. The mean age of the OSCC patients was 48.8 years and of normal buccal mucosa patients 30.3 years. In the gender distribution 70% were males and 30% were females in OSCC group and 50% male and 50% female in the normal buccal mucosa patients (Graph-1). The presence or absence of H.pylori was assessed. H.pylori was detected in the lamina propria (46.7%) and in the muscular component (10%) of OSCC tissue. Here, the p value found to be stastically significant between the OSCC and the control group (p<00.1).



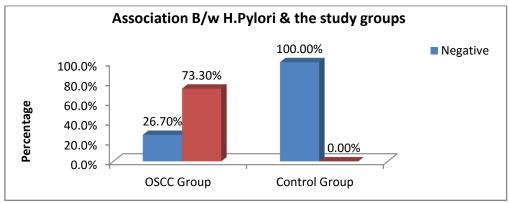
Graph 1: Shows genderwise Distribution of Study Participants among 02 groups



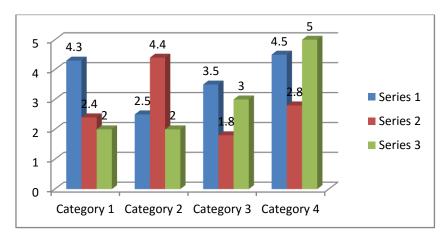
Graph 2: Agewise distribution of study Participants among 02 groups

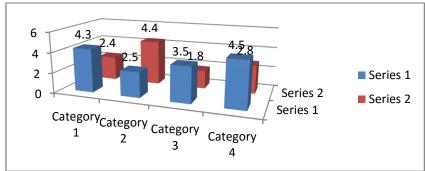
Table No. 1 Distribution of Lesion & Habit Characteristics with HP Diagnosis in OSCC group

Variables	Categories	n	%
Site of Lesion	Buccal Mucosa	30	75.0%
	Gingiva	4	10.0%
	Palate	1	2.5%
	Retro Molar Area	2	5.0%
	Tongue	3	7.5%
Habit History	Betel Quid	3	10.0%
	Tob. Chewing	13	43.3%
	Tob. Smoking	3	10.0%
	Tob. Chewing + Smoking	1	3.3%
	Info NA	10	33.3%
HP Diagnosis	Well Diff. OSCC	16	53.3%
	Mod. Diff. OSCC	11	36.7%
	Poorly. Diff. OSCC	3	10.0%

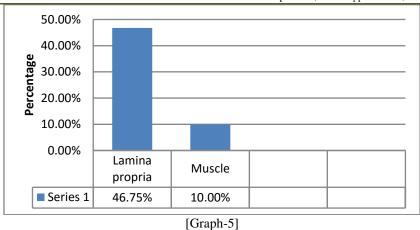


Graph no.3: Showing association B/w H.Pylori & the study groups



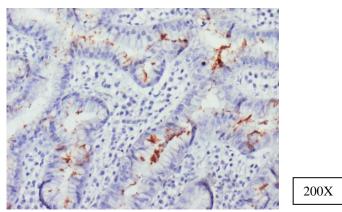


Graph no.4: Showing the presence of H.Pylori bacteria in different areas of OSCC Tumor Region

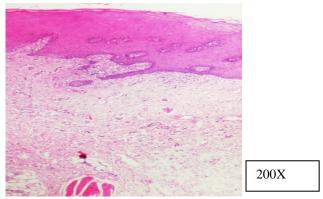


200

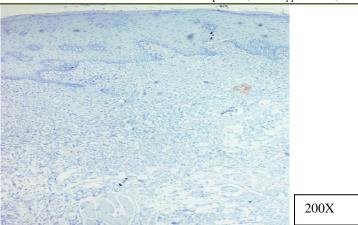
Photograph 1: Photomicrograph showing spiral shaped H.pylori seen in the lumen of gastric mucosa (H&E)



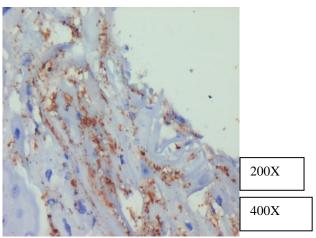
Photograph 2: Photomicrograph showing H.pylori in both spiral and coccoid form in the lumen of Gastric mucosa (IHC stain)



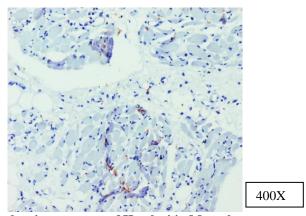
Photograph 3: Photomicrograph of normal buccal mucosa(H&E)



Photograph 4: Absence of H.pylori in Normal buccal mucosa (IHC stain)



Photograph 5: Presence of H.pylori in Lamina propria of OSCC (IHC stain).



Photograph 6: Photomicrograph showing presence of H.pylori in Muscular component of OSCC (IHC stain)

DISCUSSION

Helicobacter pylori was first discovered in the stomach of patients having gastritis and stomach ulcers in 1984 by Dr. Barry Marshall and Dr. Robin Warren and is considered to be the most common chronic bacterial infection in humans.[2] It is thought to promote tumour growth through inflammation dependent mechanism in epithelial cells leading to mucosal atrophy, inflammation and gastric malignancies.[3] In this H.pylori has gained more attention as soon as it was declared as class I

carcinogen by WHO and a definitive causative agent for gastric carcinomas.[2] Since the gate of the gastro-intestinal tract (GIT) is oral cavity; many diseases of oral cavity may affect the integrity of mucous membrane of oral cavity and the remaining portions of the GIT. It is also found to be associated with the changes in oral environment, including its microbial colonization and later infection in the oral cavity. Thus oral cavity can serve as a reservoir of micro-organisms and a source for the infection of the stomach and gut or alternatively. It also can serve as the transmission gate for external germs

which leads to further colonization of GIT.[4] As we know OSCC is the most common malignancy and accounts for almost more than 90% of all the oral malignancies and is a multifactorial disease with multiple etiological factors playing role in its pathogenesis. So we tried to correlate if any kind of association can be seen with OSCC and presence of H.pylori in the oral cavity since only few studies are listed in the literature..But there are different opinions concerning the presence of Helicobacter pylori in the oral cavity. Song Q have suggested that Helicobacter pylori may belong to the normal oral flora of the human oral cavity, maintaining a commensal relation with the host, But present in very low numbers such that the identification is very difficult.[5] There are several methods to detect H.pylori in the oral cavity like urea breadth test, culture and special stains like giemsa but in our study Immunohistochemistry (IHC) was preffered due to to its high specifity and sensitivity (>90%), to show the location of H. pylori inside the tissue (lamina propria) if present and also since it can detect coccoid /post treatment forms of H.pylori. In the present study, 30 cases of OSCC and 10 cases of normal mucosal biopsies were evaluated for H.pylori immunohistochemically. Twenty-two number of cases (73.3%) of OSCC were found to be positive for H.pylori while eight cases of OSCC and all the 10 cases of normal buccal mucosa were negative for H.pylori. H. pylori has been detected in various regions of the oral cavity and a high variation is seen in its presence in the oral cavity i.e dental plaque (82.3%), gargles (51.1%) and the dorsum surface of tongue (37.5%).[6] The two possible mechanisms involved in H. pylori pathogenesis are firstly, H. pylori interacts with surface epithelial cells, developing direct cell damage or producing proinflammatory mediators.[7-9] Secondly, H. pylori reaches the underlying connective tissue to stimulate an immune response, leading to the release of various cytokines and oxygen radicals that transform the chronic gastritis into gastroduodenal ulcers and gastric carcinoma. Also H. pylori produce some extracellular products that cause local and systemic immune

Responses, which can result in tissue damage. [10-12] With the advancement of biochemical techniques, new information about the pathogenicity and virulence factors of H.pylori has emerged, indicating that infection by H.pylori requires a complex interaction of both bacterial and host factors. Bacterial proteins like urease are necessary for colonization of gastric mucosa by H.pylori. The urease produced by the bacteria alters the microenvironment of the organism to facilitate colonization. The bacteria adhers to the surface of gastric mucosa due to its mobility by using its flagella and attaches via adhesions to glycolipid receptor on the apical membrane of surface epithelial cells. H.pylori produces cecropins to inhibit the growth of competing organisms. Once attached to gastric mucosa, H. pylori cause tissue injury by a complex cascade of events that depends on both the organism and the host. H. pylori,

like all gram-negative bacteria have in its cell wall lipopolysaccharide, which acts to disrupt mucosal integrity. Furthermore, H.pylori releases several pathogenic proteins that induce cell injury. Once it gets colonized in the gastric mucosa, the immunogenic properties of H.pylori induce an inflammatory reaction with neutrophilic gastritis that ultimately results.

In the clinical manifestations of the infection.[13] In present study the mean age of occurrence of OSCC was found to be 48.8 or middle age (Graph-2) same as given in the literature, suggestive of either late development of OSCC or due to late detection of OSCC because many of the times it get unnoticed when it is asymptomatic. Male predominance is seen in our study (Graph -1) with almost 70% being male and 30% female and this can be attributed to the more involvement of male towards using various forms of tobacco and also the highest percentage (43.3%) is found in the patients who use chewable form of tobacco as compared to other forms of tobacco. This also suggest that why in our study buccal mucosa (70%) was the most common site involved due to tobacco chewing habit of the individuals in Indian subcontinent. All the three demographic data i.e age of occurrence, sex and site involved are in accordance with earlier studies. [14-16] Most of the cases reported are well differentiated. followed by moderately and poorly differentiated OSCC (Table no.1). In our present study, 22 (73.3 %) cases of OSCCs showed H. pylori positivity (Graph no.3). Fernando et al done a study to show the presence of H.pylori in oral cancer patients who are betel and nonbetel chewers and found that as compared to non-betel chewers the H.pylori presence was statistically significan in betel chewers.¹⁷Rubin et al, working on 61 samples from head and neck malignant and premalignant conditions, detected H. pylori positivity in 16.3% of oral cavity samples.[18] Grimm et al in 2014 analyzed the prevelance and influence of H.pylori in OSCC immunohistochemically and found that H.pylori prevelance was found to be 21.5% and was associated with the disease free survival.[2] Dayamma et al in found positive result in only 3% of cases of OSCC using culture and PCR technique but the male predominance and age involvement coincides with our study.[21]Okuda et al in 2000 done a study using swab samples of the oral mucosa and cancer lesion surfaces, but no positive PCR results were obtained.[19] Chitsazi et al in 2007 reported that 40% of 39 patients had viable H. pylori in their oral cavities despite H. pylori eradication also 56% of those without detectable H. pylori in the mouth before treatment had H. pylori in the oral cavity when reexamined after H. pylori eradication.[20]In the present study, H. pylori oral colonization was seen in both the coccoid and the spiral forms. Wang et al suggested that the coccoid form of H. pylori is viable and maintains the integrity of the nucleic acid contents, involved in active protein synthesis and is able to synthesize DNA. The present study detected the coccoid form of H. pylori, which might be a proof for its long-standing presence in the oral cavity and revealing the role of H. pylori in the pathogenesis of OSCC. Several studies support the hypothesis that the oral cavity is a reservoir for reinfection of the stomach.[22] On the other hand, some other investigations have shown that presence of H. pylori in the oral cavity does not relate to gastric infection and that H. pylori can also be found in the oral cavity without any gastric infection.[23-25] Song et al, shown that H. pylori DNA sequences differed between oral samples and gastric samples within the same individual.[5]Previous studies on the gastric mucosa indicated the presence of H. pylori in the lamina propria, the intercellular space as well as in the gastric lumen [24] coinciding with our study where we have seen presence of H.pylori in the lamina propria. The presence of H. pylori in the stromal cell of the lamina propria, far from basement membrane, the epithelial invasion.[25] As seen in our study 14 cases of OSCC (46.7%) were positive for the lamina propria (Graph no.4). Various studies have shown H. pylori invasion into the lamina propria of gastric mucosa, which can be an important factor in the induction and development of gastric inflammation.[26-27] In the present study H. pylori was found in the lamina propria in 46.7% and muscle (10%) which can be clear evidence for the invasion of the bacteria..In our study the detection of H.pylori is found highest in lamina propria suggestive of the invasive behaviour of the bacteria especially in ulcerated cases where they get more serum factors.Our results are very much similar to the study done by Soussan irani et al where H.pylori positivity is seen in 26.5% cases of OSCC but here H.pylori presence is more as we have taken more number of ulcerated cases, which are devoid of epithelium.

CONCLUSION

Hence, we can suggest that there is a positive relationship between the presence of H.pylori in the oral cavity and OSCC and it can act as risk factor for the development of OSCC. Since, it causes direct cell damage by producing various proinflammatory mediators and also by stimulating an immune response, can lead to production of different cytokines and oxygen radicals which might help in further progression of OSCC. [28-29]

However, We also assessed for the presence of H.pylori in normal mucosal biopsies. None of the buccal mucosal biopsies in our study showed positivity for H.pylori. So this raises the doubt whether oral cavity acts as a real H.pylori reservoir or whether it is transiently stored in mouth as a result of gastroesophageal reflux or when passing to the stomach.³¹ Before concluding, several limitations of our current study might be considered as follows. Firstly, we failed to obtain clinical history of the patients including gastrointestinal symptoms or other stomach disorders. Secondly it is always advisable to do two or more diagnostic methods to confirm the presence of H.pylori. Thirdly the presence

of H.pylori in oral cavity is low in number and may be suppressed by the complex oral microflora.[32] thus further large-scale research with appropriate diagnostic methods in this field could open doors to understand the true nature of H.pylori in OSCC.

BIBLIOGRAPHY

- 1. Zou Q H, Li R Q, Helicobacter pylori in the oral cavity and gastric mucosa: a Meta analysis. J Oral Pathol Med 2011; 40: 317-324.
- Grimm M, Munz A, Exarchow A, Polligkeit J, Reinert S. Immunohistochemical detection of Helicobacter pylori without association of TLR5 expression in oral squamous cell carcinoma. J Oral Pathol Med 2014; 43:35-44.
- 3. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell 2011; 144: 646–74.
- 4. Hardin F J, Wright R A. Helicobacter pylori: Review and Update. Hospital physician may2012; 23-31.
- Song Q, Lange T, Spahr A, Adler G, Bode G. Characteristic distribution pattern of Helicobacter pylori in dental plaque and saliva detected with nested PCR. J Med Microbiol 2000; 49:349-53.
- 6. Gao J, Li Y, Wang Q, Qi C, Zhu S. Correlation between dis-tribution of Helicobacter pylori in oral cavity and chronic stomach conditions. *J Huazhong Univ Sci Technolog Med Sci* 2011; 31:409-12.
- Amieva MR, Vogelmann R, Covacci A, Tompkins LS, Nel-son WJ, Falkow S. Disruption of the epithelial apical-junctional complex by Helicobacter pylori Cag A. Science 2003;300:1430-4.
- 8. Naumann M, Crabtree JE. Helicobacter pyloriinduced epithelial cell signalling in gastric carcinogenesis. *Trends Microbiol* 2004; 12:29-36.
- 9. Peek RM Jr. IV. Helicobacter pylori strain-specific activa-tion of signal transduction cascades related to gastric in-flammation. *Am J Physiol Gastrointest Liver Physiol* 2001;280: G525-30.
- Amieva MR, Vogelmann R, Covacci A, Tompkins LS, Nel-son WJ, Falkow S. Disruption of the epithelial apical-junctional complex by Helicobacter pylori Cag A. Science 2003;300:1430-4.
- 11. Naumann M, Crabtree JE. Helicobacter pyloriinduced epithelial cell signalling in gastric carcinogenesis. *Trends Microbiol* 2004; 12:29-36.
- 12. Peek RM Jr. IV. Helicobacter pylori strain-specific activa-tion of signal transduction cascades related to gastric in-flammation. *Am J Physiol Gastrointest Liver Physiol* 2001;280: G525-30.
- 13. Panahi O, Rezaei S, Marzi M, Asghari FS. Helicobacter pylori and oral cavity inflammation. JPCS 2011; 2:13-5.
- Sudhakar U, Anusuya CN, Ramakrishnan T, Vijayalakshmi R. Isolation of Helicobacter pylori from dental plaque: A microbiological study. *J Indian Soc Periodontol* 2008; 12:67-72.
- 15. Pires F R, Ramos A B, Oliveira J B C, Tavares A S, LuzP S R, Santos T C R B. Oral squamous cell

- carcinoma: clinicopathological features from 346 cases from a single Oral Pathology service during an 8-year period. *J Appl Oral Sci.*, 2013;21(5):460-763. (well differentiated): A case report. *Journal of Dentistry and Oral Hygiene*, 2013; 5(4): 31-34.
- MarocchioL S, Lima J, SperandioF F, CorrêaL and de Sousa S O M. Oral squamous cell carcinoma: an analysis of 1,564 cases showing advances in early detection. *Journal of Oral Science*, 2010; 52 (2): 267-273.
- 17. Fernando N, Jayakumar G, Perera N, Amarasingha I, Meedin F, Holton J. Presence of *Helicobacter pylori* in betel chewers and non-betel chewers with and without oral cancers. BMC Oral Health 2009; 9:23
- 18. Rubin JS, Benjamin E, Prior A, Lavy J. The prevalence of Helicobacter pylori infection in malignant and premalignant conditions of the head and neck. *J Laryngol Otol* 2003; 117:118-21.
- 19. Okuda K, Ishihara K, Miura T, *et al.*, Helicobacter pylori may have only a transient presence in the oral cavity and on the surface of oral cancer. Microbiol Immunol 2000; 44: 385–8.
- Chitsazi MT, Fattahi E, Farahani RM, FattahiS. Helicobacter pylori in the dental plaque: Is it of diagnostic value for gas-tric infection? *Med Oral Patol Oral Cir Bucal* 2006;11: E325-8.
- 21. Anand D, Vineeta S, Mridula S, Royana S, Manoj P. Helicobacter pylori and oral cancer: possible association in a preliminary case control study. Asian Pac J Cancer Prev 2011; 12:1333-36.
- 22. Dowsett SA, Kowolik MJ. Oral Helicobacter pylori: Can we stomach it? *Crit Rev Oral Biol Med* 2003; 14:226-33.
- 23. Oshowo A, Gillam D, Botha A, Tunio M, Holton J, Bou-los P, *et al.*, Helicobacter pylori: The mouth,

- stomach, and gut axis. *Ann Periodontol* 1998; 3:276-80.
- 24. Teoman I, Ozmeriç N, Ozcan G, Alaaddinoğlu E, Dumlu S, Akyön Y, *et al.*, Comparison of different methods to detect Helicobacter pylori in the dental plaque of dyspeptic pa-tients. *Clin Oral Investig* 2007; 11:201-5.
- Necchi V, Candusso ME, Tava F, Luinetti O, Ventura U, Fiocca R, et al., Intracellular, intercellular, and stromal inva-sion of gastric mucosa, preneoplastic lesions, and cancer by Helicobacter pylori. Gastroenterology 2007; 132:1009-23.
- Ito T, Kobayashi D, Uchida K, Takemura T, Nagaoka S, Kobayashi I, et al., Helicobacter pylori invade the gastric mucosa and translocates to the gastric lymph nodes. Lab In-vest 2008; 88:664-81.
- 27. Peek RM Jr. IV. Helicobacter pylori strain-specific activa-tion of signal transduction cascades related to gastric in-flammation. *Am J Physiol Gastrointest Liver Physiol* 2001;280: G525-30.
- 28. Ndawula EM, Owen RJ, Mihr G, Borman P, Hurtado A. Helicobacter pylori bacteraemia. *Eur J Clin Microbiol Infect Dis* 1994; 13:621.
- Rennemo E, Zatterstrom U, Boysen M. Impact of second primary tumors on survival in head and neck cancer: An analysis of 2,063 cases. *Laryngoscope* 2008; 118:1350-6.
- 30. Mc Carthy P L, Shklar G, editors. Lichen planus. Diseases of the oral mucosa.2nd ed. Philadelphia: Lea and Febriger; 1980: 203-204.
- 31. Burgers R, Schneider-Brachert W, Reischl U, Behr A, Hiller K-A, Lehn N, *et al.*, Helicobacter pylori in human oral cavity and stomach. Eur J Oral Sci 2008; 116:297–304.